

GENESIS
Generalized-ensemble simulation system



マルチスケール分子動力学ソフトウェアGENESISの並列化

Supercomputing Japan 2024
2024/03/12 タワーホール船堀

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RIKEN Center for Biosystems Dynamics Research
RIKEN Cluster for Pioneering Research
RIKEN iTHEMS

BDR



Glyco-lipidologue



BIE

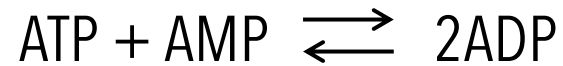


GROSS BIOLOGY

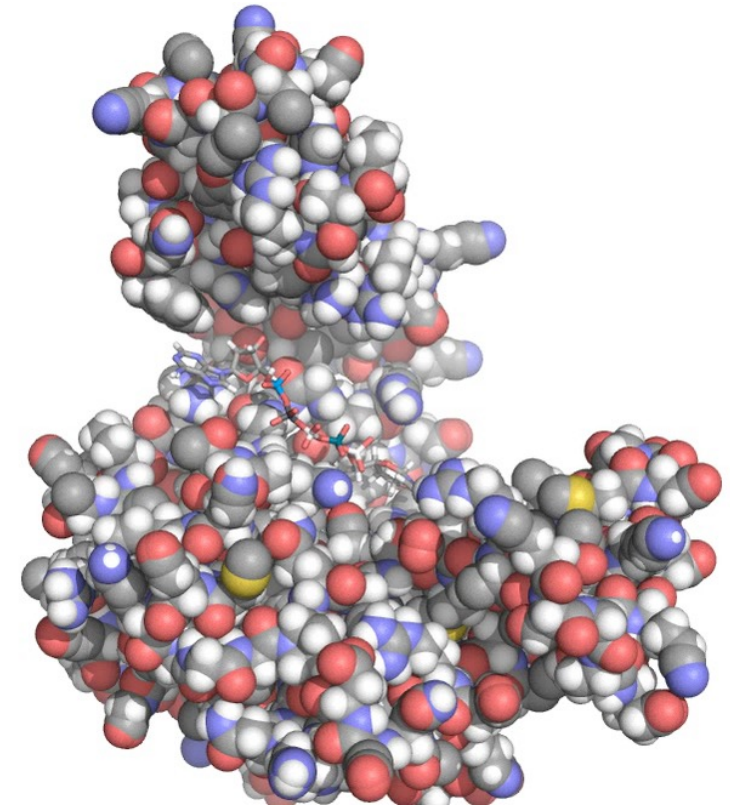
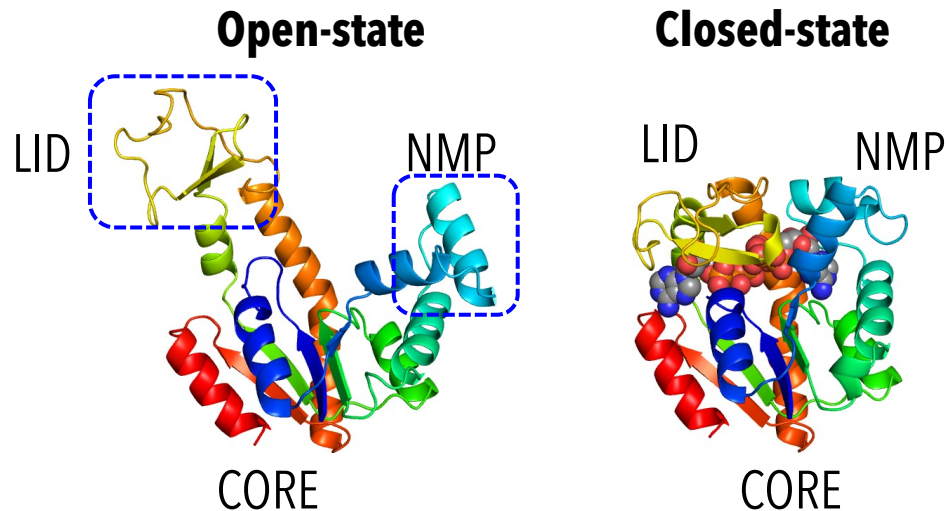
Protein Dynamics and Functions

- Protein structures are determined by X-ray crystallography, NMR , and cryo-EM at atomic resolutions.
- Proteins are flexible for their functions in a living cell .

Enzyme reaction by Adenylate Kinase (Adk)



Conformational changes of Adk

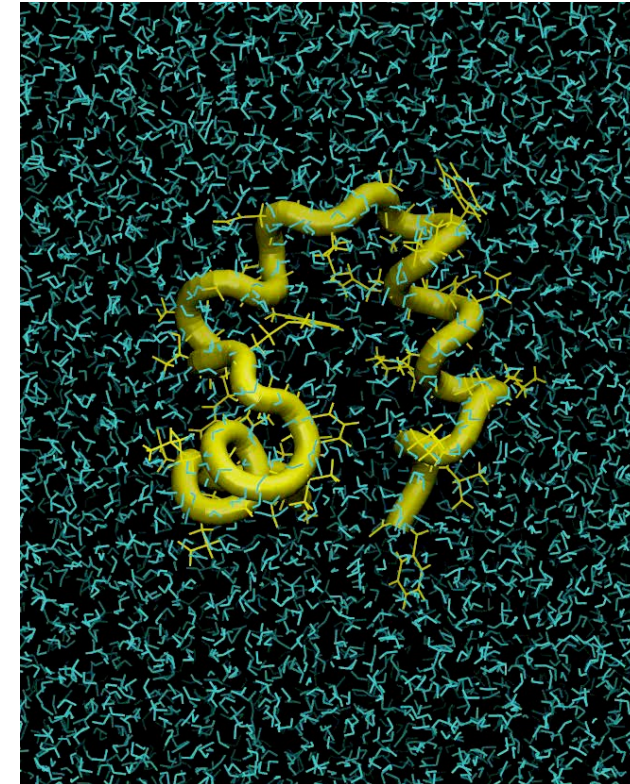


Molecular dynamics (MD) simulation

- MD is useful to predict molecular motions at the atomic resolutions.
- In classical MD, newton's equation of motion is solved iteratively.

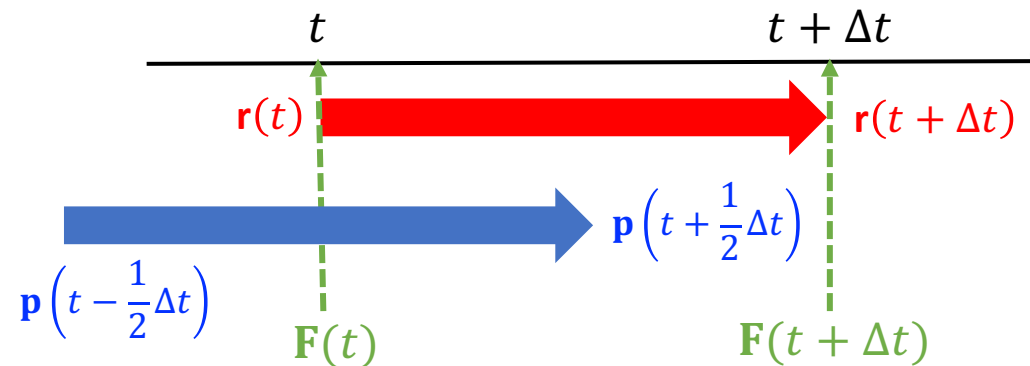
$$F = ma$$

- Time step Δt should be very short to reproduce fast vibrations in molecules. (10^{-15} sec). Many iterations are required to detect protein dynamics.



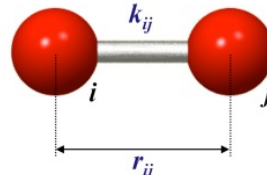
$$\mathbf{r}_i(t + \Delta t) = \mathbf{r}_i(t) + \frac{\mathbf{p}_i\left(t + \frac{1}{2}\Delta t\right) \Delta t}{m_i}$$

$$\mathbf{p}_i\left(t + \frac{1}{2}\Delta t\right) = \mathbf{p}_i\left(t - \frac{1}{2}\Delta t\right) + \mathbf{F}_i(t)\Delta t$$



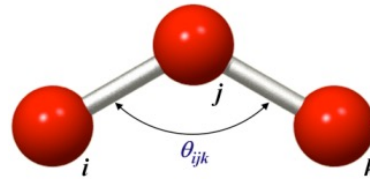
Molecular Force Field Function

$$V(\mathbf{r}) = \sum_{\text{bonds}} k_b (r_{i,i+1} - r_0)^2$$



Bond term

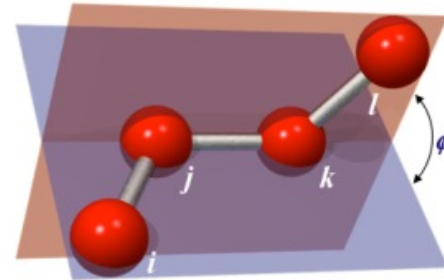
$$+ \sum_{\text{angles}} k_a (\theta_{i,i+1,i+2} - \theta_0)^2$$



Angle term

taken from PumMa HP

$$+ \sum_{\text{torsions}} k_t \left[1 + \cos(n\phi_{i,i+1,i+2,i+3} - \omega) \right]$$



Dihedral term

$$+ \sum_i \sum_{j>i} \epsilon \left[\left(\frac{\sigma}{r_{ij}} \right)^{12} - \left(\frac{\sigma}{r_{ij}} \right)^6 \right]$$

vdW term
(solved with cutoff approximation)

$$+ \sum_i \sum_{j>i} \frac{q_i q_j}{4\pi\epsilon_0 r_{ij}}$$

Electrostatic term

$O(N^2)$

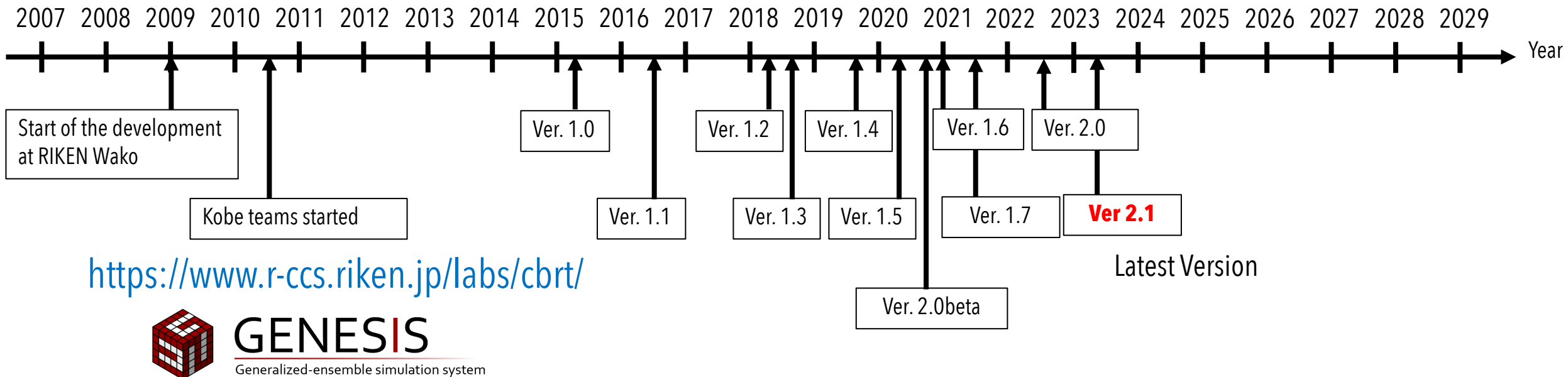
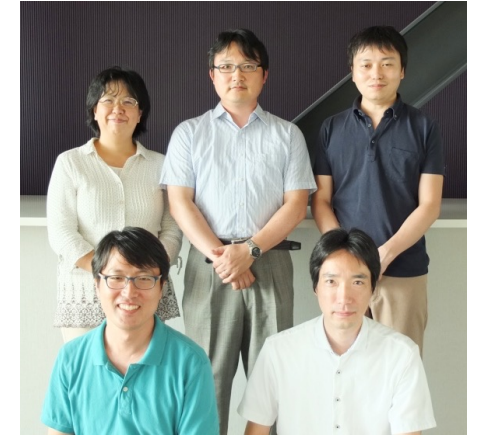
CHARMM, AMBER, OPLS, and other Force Fields are available for various molecules.



Development of GENESIS

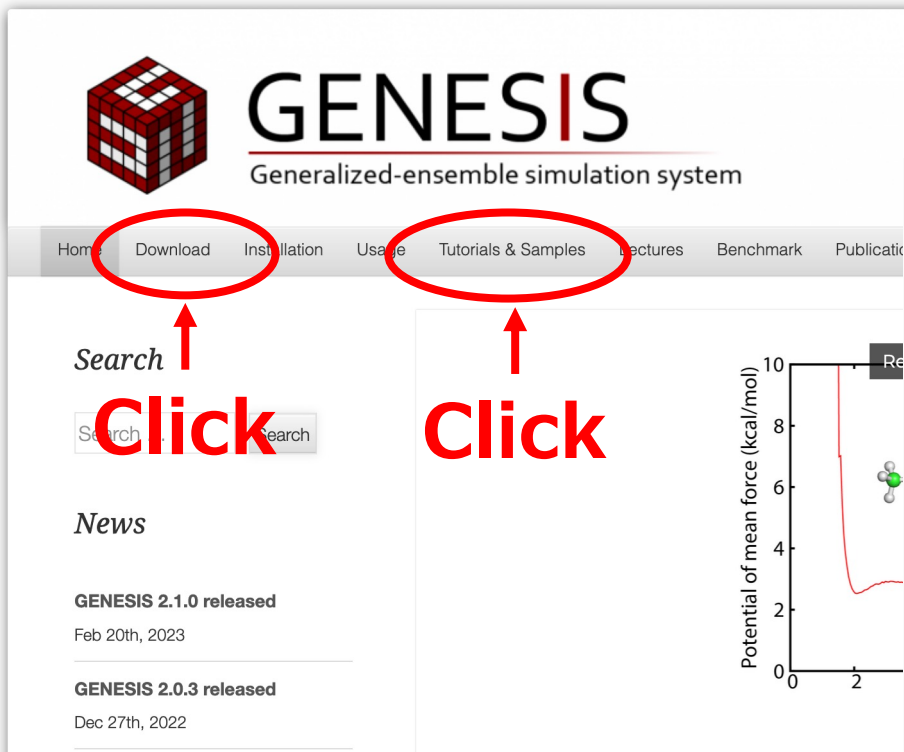
- We have developed GENESIS software since 2019.
- GENESIS can show good performances on K and Fugaku.
- GENESIS is freely available for everyone under LGPLv3 license.

Initial Developers

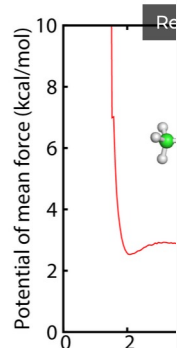


GENESIS is freely available

<https://www.r-ccs.riken.jp/labs/cbrt/>



The screenshot shows the GENESIS website home page. The navigation menu includes Home, Download, Installation, Usage, Tutorials & Samples, Lectures, Benchmark, and Publications. The 'Download' and 'Tutorials & Samples' links are circled in red, with red arrows pointing to them and the word 'Click' written in large red text. A search bar is located below the navigation menu. The 'News' section features two entries: 'GENESIS 2.1.0 released' (Feb 20th, 2023) and 'GENESIS 2.0.3 released' (Dec 27th, 2022).



The screenshot shows the 'Tutorials 2022' page on the GENESIS website. The navigation menu includes Home, Download, Installation, Usage, Tutorials & Samples, Lectures, Benchmark, Publications, Links, and About. The 'Tutorials 2022' page content includes a search bar, a news section with three entries: 'GENESIS 2.1.0 released' (Feb 20th, 2023), 'GENESIS 2.0.3 released' (Dec 27th, 2022), and 'GENESIS 2.0.2 released' (Dec 20th, 2022), and a main text area titled 'Tutorials 2022' which provides instructions for using GENESIS version 2.0, including recommendations to install VMD and gnuplot, and to study the Level 1 Basic tutorials first.

How can we accelerate MD simulations to understand the slow conformational changes of proteins (if we can use supercomputers) ?

The Supercomputer K



Node	CPU	SPARC64™ VIIIfx 2GHz
	Performance	128 GF (16 GF x 8 cores)
	Memory	16 GB
Number of Nodes per Rack		864
Total Number of Nodes		82,944
Network		Tofu Interconnect (6D Mesh/Torus)
Peak Performance		10.62 PF
Total Memory		1.26 PB
File IO		Fujitsu Exabyte File System (FEFS)
Storage		30 PB

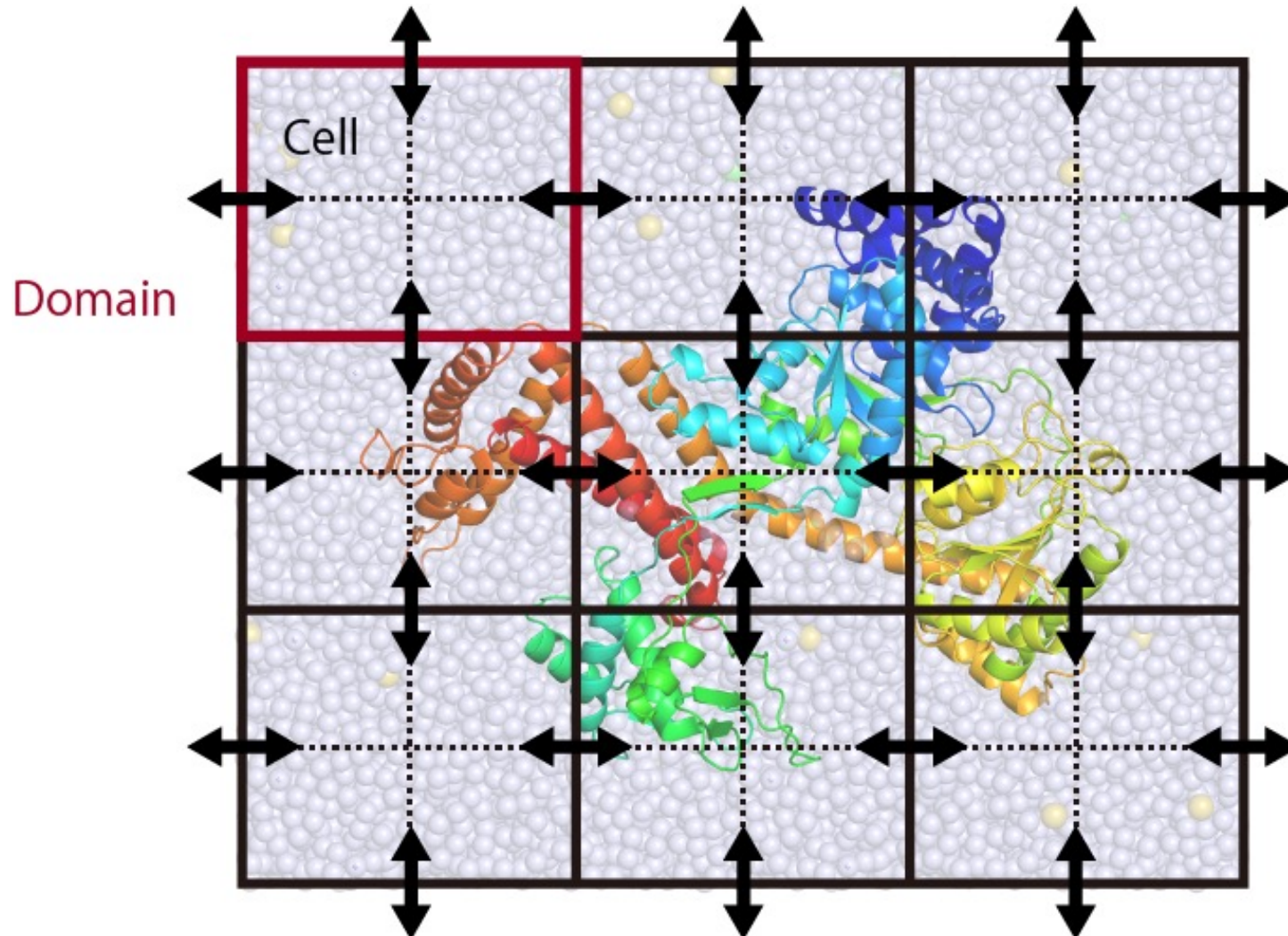


Top500	No.1 for 2 consecutive times since Jun. 2011
Graph500	No.1 in June 2014 & for 6 consecutive times since Jul. 2015
HPCG	No.2 for 4 consecutive times since Nov. 2014 No.1 for 3 consecutive times since Nov. 2016

The K computer strikes a balance between performance in calculation, memory, and communication.

Shared use of the K computer has ended on August 16, 2019.

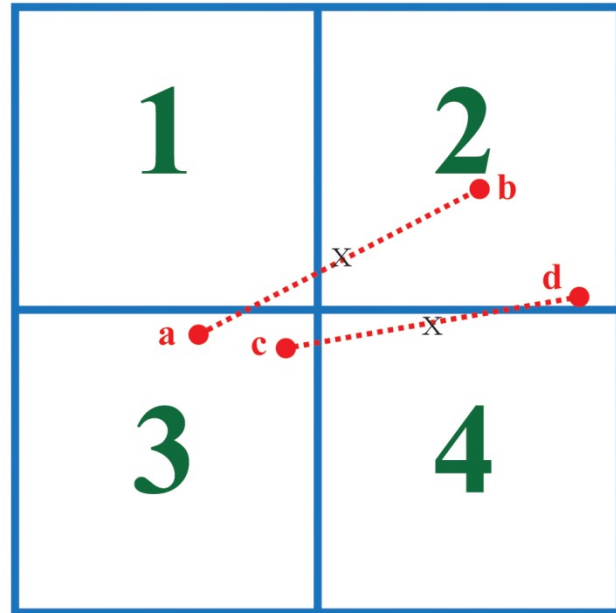
Domain Decomposition Scheme



Divide a whole space into subdomains and cells → Greatly reduce the memory

Midpoint Cell Method

(a)



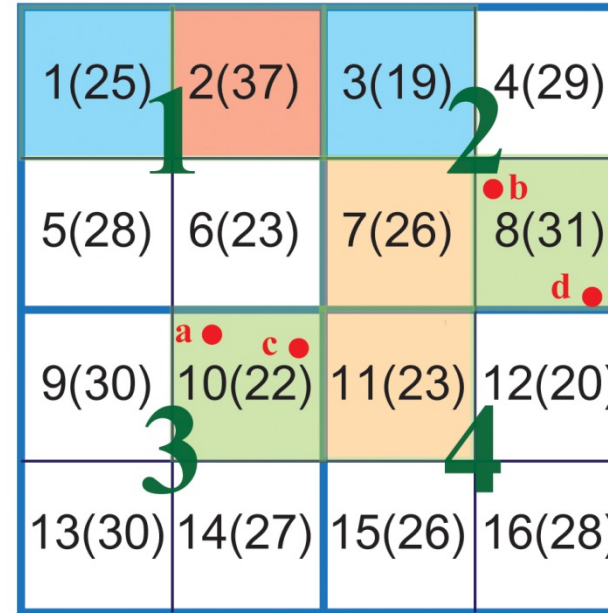
Midpoint Method:

Interaction is computed at the cell where the midpoint of two interacting particles is located.

(○) Low communication cost

(X) Need to decide the midpoint every time step.

(b)



Midpoint Cell Method:

Interaction is computed at the midpoint cell for each cell pair.

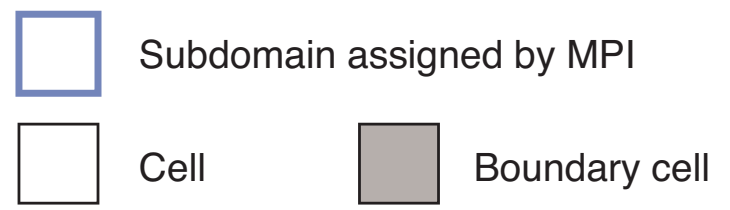
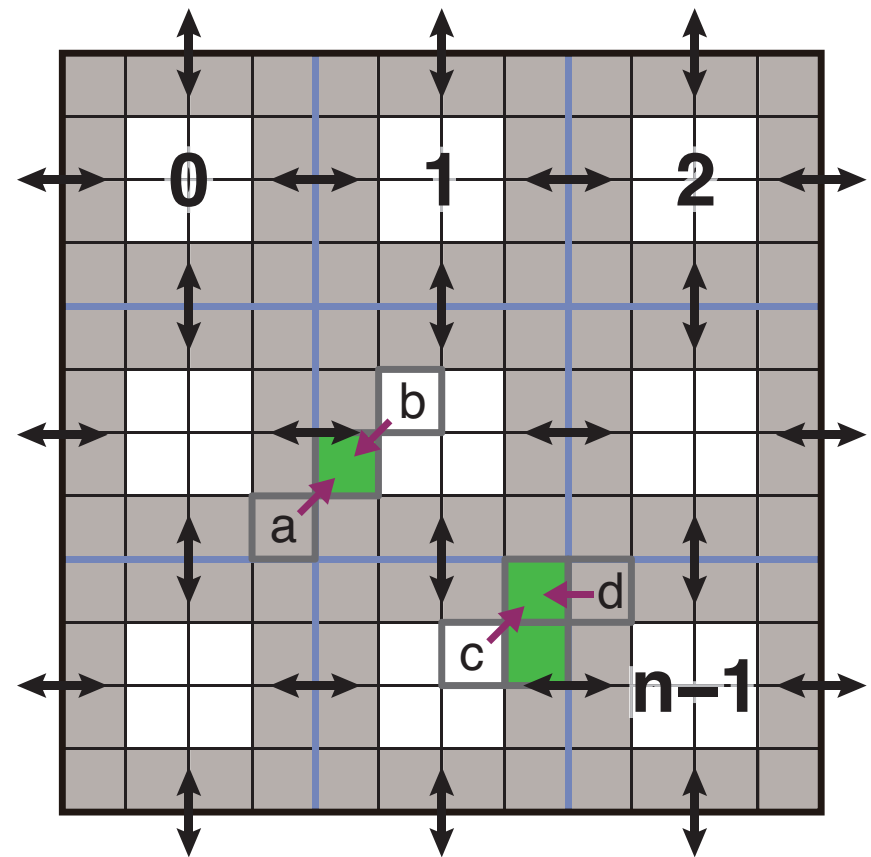
(○) Low communication cost

(○) Midpoint cell is decided only once.



Subdomains and cells

(a)



1. The basic domain decomposition scheme is the midpoint cell method.
2. A whole system is divided into **subdomains** and **cells**.
3. Information of the particles residing at each **subdomain** (plus surrounding cells) is stored at each **MPI process**.
4. Each subdomain is further divided into **cells** for **OpenMP thread** parallelization.

This method is suitable to Hybrid parallelization.

Intracellular Environments

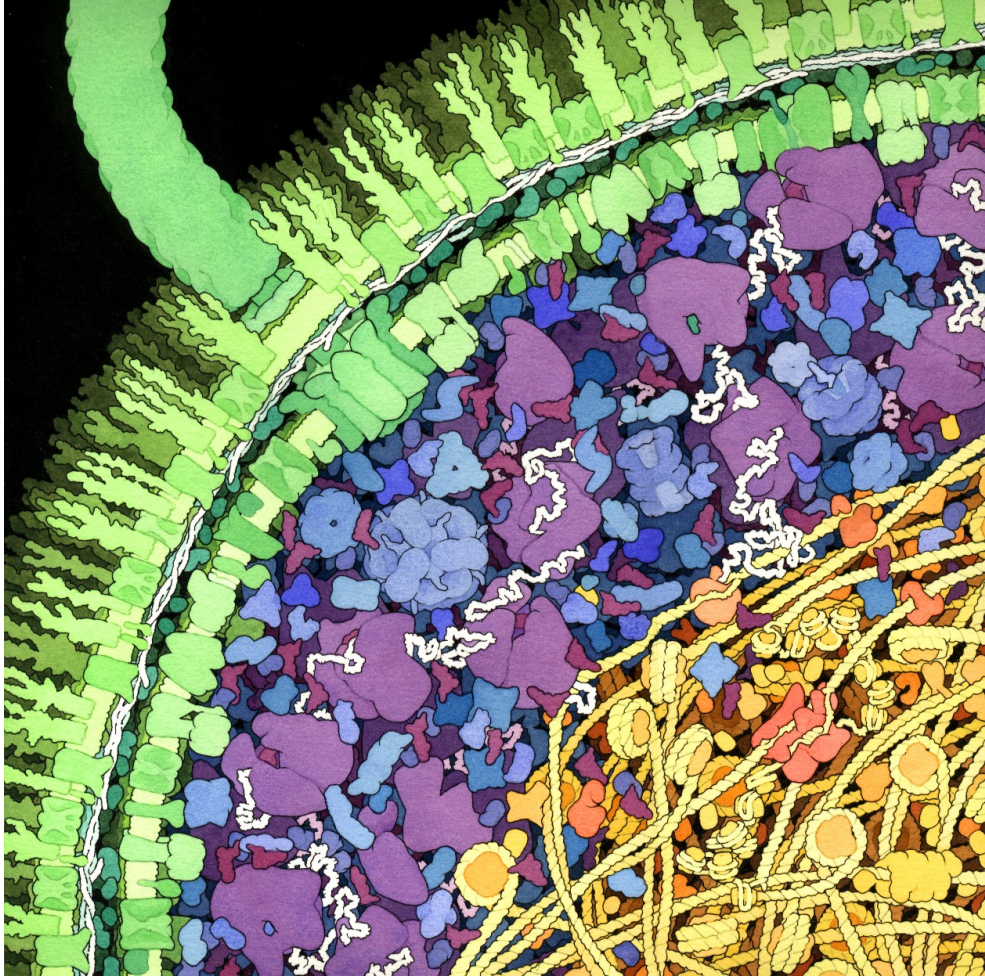


Illustration by Goodsell for E. coli cell

- Concentration of proteins and RNA in the E. coli cell is 300-400mg/ml.
- 20-40% of the volume inside of the cell is occupied by macromolecules.
- This condition has been called as **macromolecular crowding**, which can affect structure, dynamics, and functions of macromolecules in the cell.

Zimmerman and Minton, *Ann. Rev. Biophys. Biomol. Struct.* (1993) 22: 27-65.

Zhou, H.X., Rivas, G., Minton, A.G., *Ann. Rev. Biophys.* (2008) 37, 375-397.

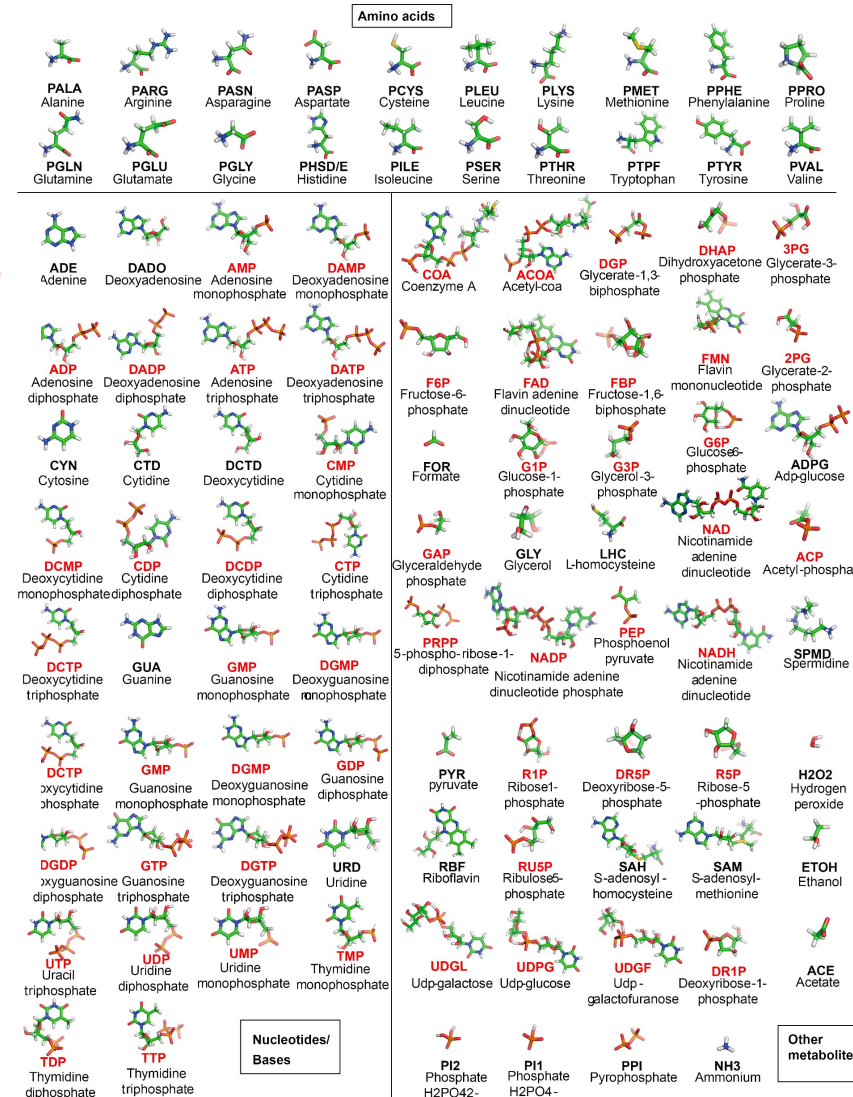
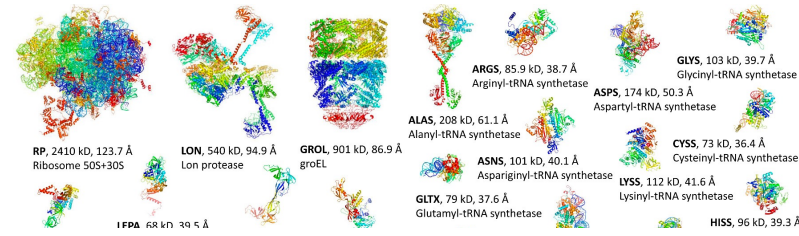


Atomistic Modeling of the Cytoplasm

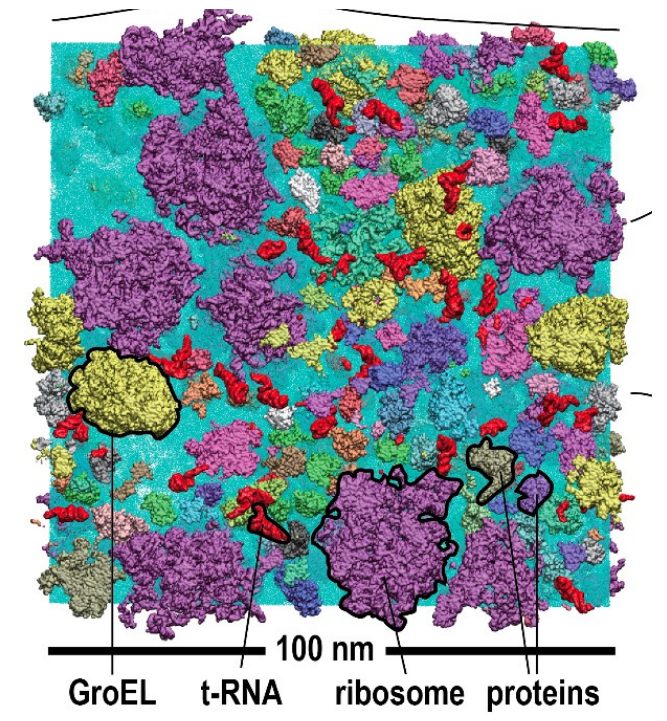
Homology models of protein structures

Metabolites in *M. Genitalium*

Fully hydrated models



- # of atoms: 103,708,785
- # of proteins: 1238
- # of RNAs: 284
- # of metabolites: 41,006
- # of ions: 214,000
- # of water: 26,263,505



AlphaFold and RosettaFold were not available in 2015.

M. Feig
(Michigan State University)

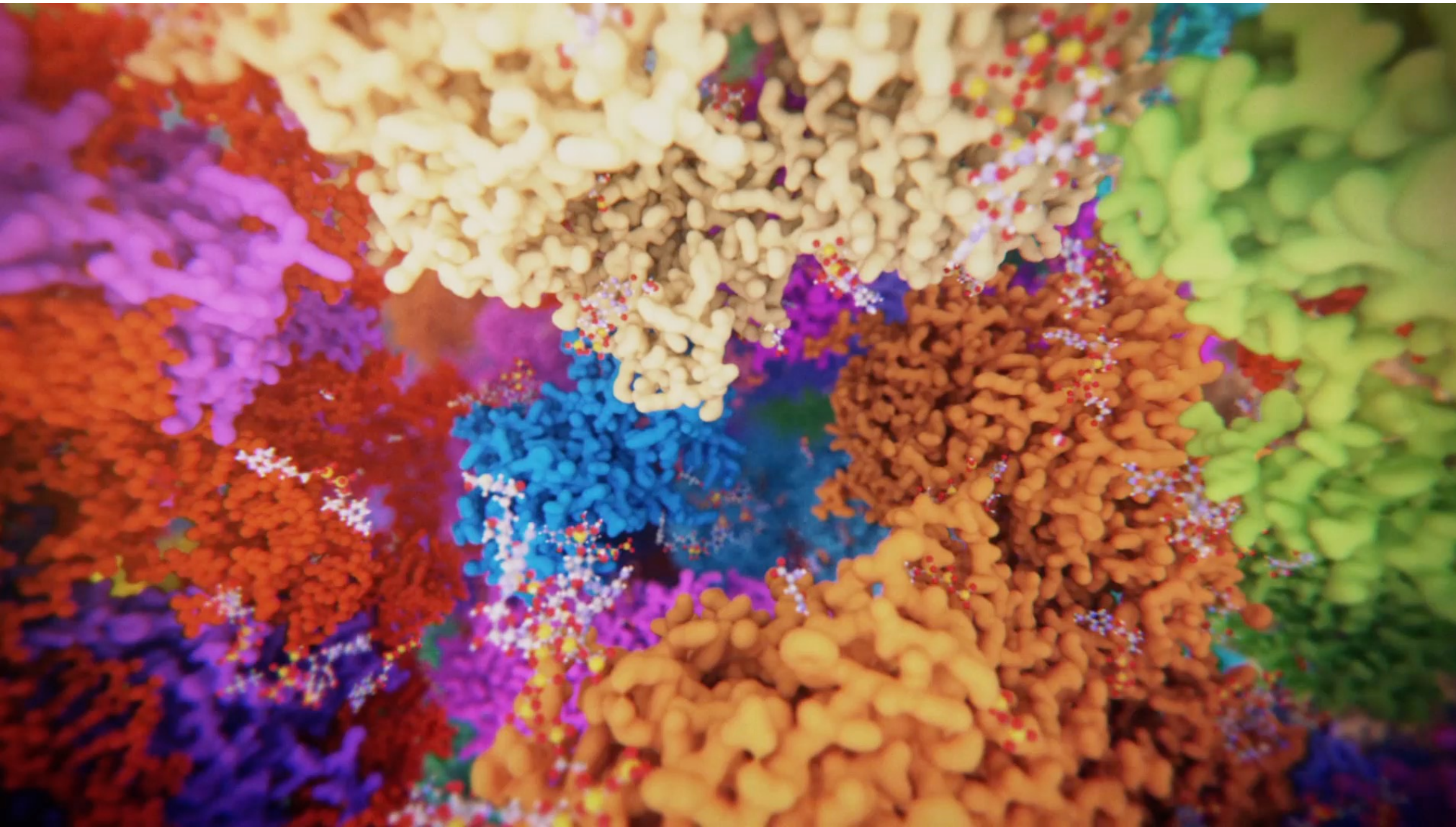
Trigger factor FRR, 22 kD, 25.0 Å
Ribosome recycling factor UNAJ, 202 kD, 61.6 Å
MRNA, 33 kD, 33.4 Å mRNA
PEPA, 295 kD, 56.4 Å
Cytosol aminopeptidase RNR, 87 kD, 3 Å Ribonuclease
TMK, 48 kD, 30.9 Å
Thymidylate kinase DEOD, 159 kD, 44.1 Å
Purine nucleoside phosphorylase
UPP, 111 kD, 39.6 Å
Uracil phosphoryltransferase CDD, 60 kD, 30.8 Å
Cytidine deaminase
APT, 40 kD, 28.0 Å
Adenine CMK, 25 kD, 24.4 Å
Cytidylate kinase
GMK, 55 kD, 36.1 Å
Guanylate kinase PYRH, 163 kD, 46.4 Å
Uridylate kinase
ACKA, 89 kD, 38.2 Å
Acetate kinase GAPA, 146 kD, 42.7 Å
Glyceraldehyde 3-phosphate
PYK, 229 kD, 54.6 Å
Pyruvate kinase
NAMP, 104 kD, 38.8 Å
Nicotinamide phosphoryltransferase

Disordered regions were not included in the models.

Nucleotidase PRS, 78 kD, 40 Å
Ribose phosphate pyrophosphokinase
YID1, 33 kD, 25.6 Å
Sugar phosphatase
GLPK, 228 kD, 51.5 Å
Glycerol kinase
OHBR, 31 kD, 26.3 Å
organic hydroperoxide resistant protein
RPIB, 34 kD, 25.8 Å
Ribose 5-phosphate isomerase B
CHOK, 33 kD, 27.5 Å
Choline kinase
GALU, 130 kD, 43.3 Å
UTP-glucose-1-phosphate uridylyltransferase
PPNK, 117 kD, 42.4 Å
Inorganic polyphosphate kinase
PPA, 65 kD, 36.1 Å
Inorganic pyrophosphatase
COF, 33 kD, 25.3 Å
Hydrolase
FRED, 19 kD, 22.1 Å
Flavin mononucleotide reductase
THU, 42 kD, 27.6 Å
Thiamine kinase
FOLD, 90 kD, 35.7 Å
Methylenetetrahydrofolate dehydrogenase
PTA, 71 kD, 35.5 Å
Phosphate acetyltransferase
NOX, 51 kD, 31.7 Å
NADH oxidase
RIFB, 62 kD, 35.1 Å
Riboflavin kinase
Pyruvate dehydrogenase
METK, 85 kD, 35.3 Å
S-adenosylmethionine synthase
PGM, 57 kD, 30.9 Å
Phosphoglucomutase
MTHF, 19 kD, 21.7 Å
5-formyltetrahydrofolate cycloligase
UMP, Uridine monophosphate
TMP, Thymidine monophosphate
TTP, Thymidine triphosphate



The cytoplasm in *M. genitalium*



The first all-atom MD calculation of the bacterial cytoplasm (> 100 M atoms)



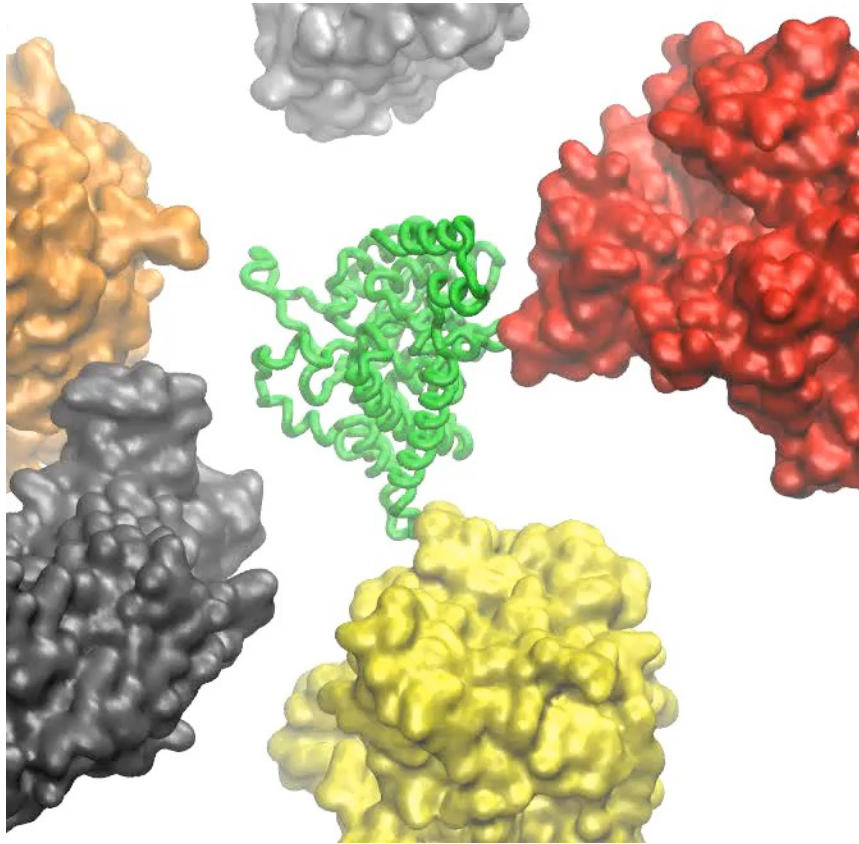
Isseki Yu (RIKEN
→ Maebashi Inst. Tech.)



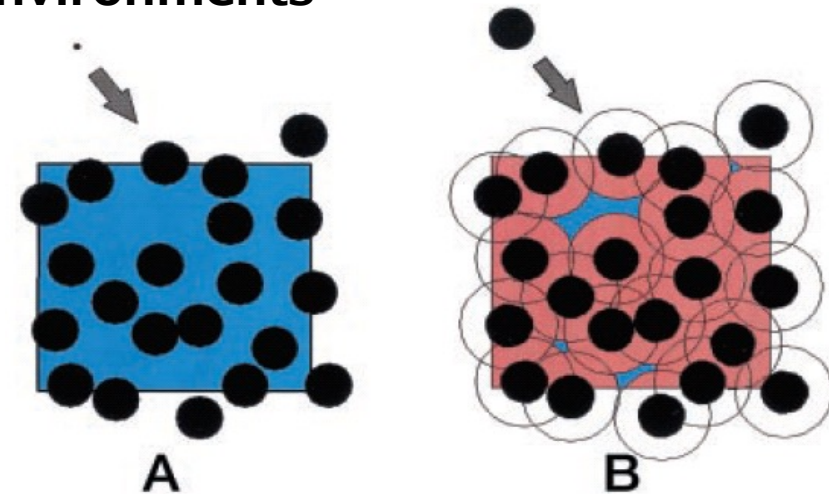
M. Feig
(Michigan State University)

Destabilization in Crowded Environments

PDHA destabilization in Crowded Environments



Volume Exclusion Effect in crowded environments



Blue: available, Red: excluded (Minton, 2001)

Free space for large molecules is very limited.

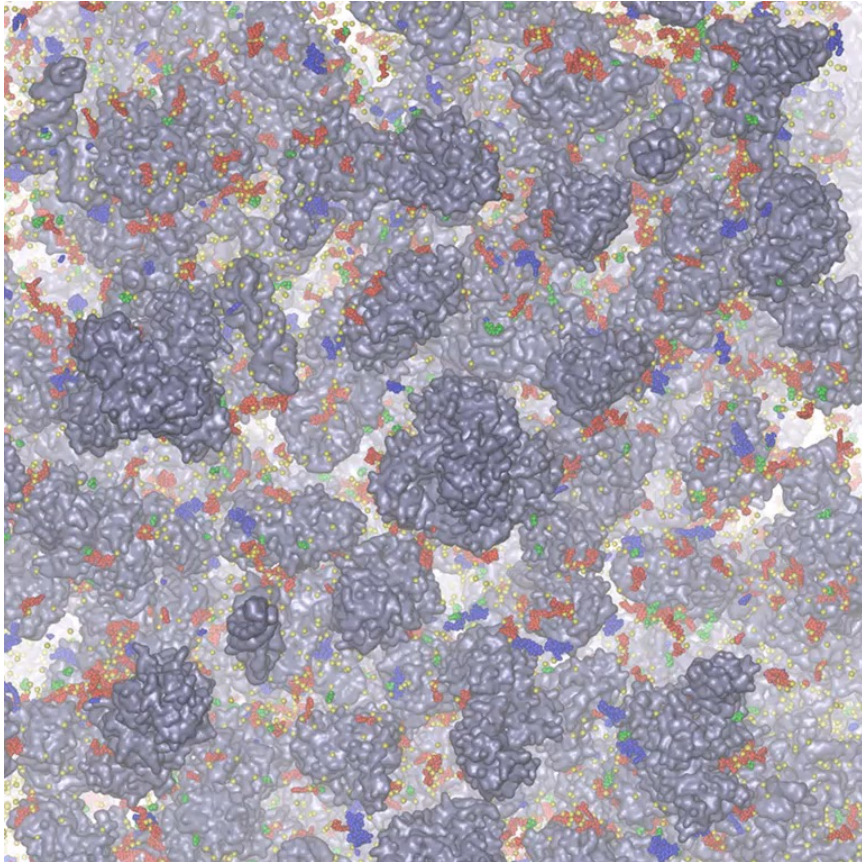


In the crowded condition, compact structures are preferred.

This is inconsistent with the excluded volume effect.

ATP controls solubility

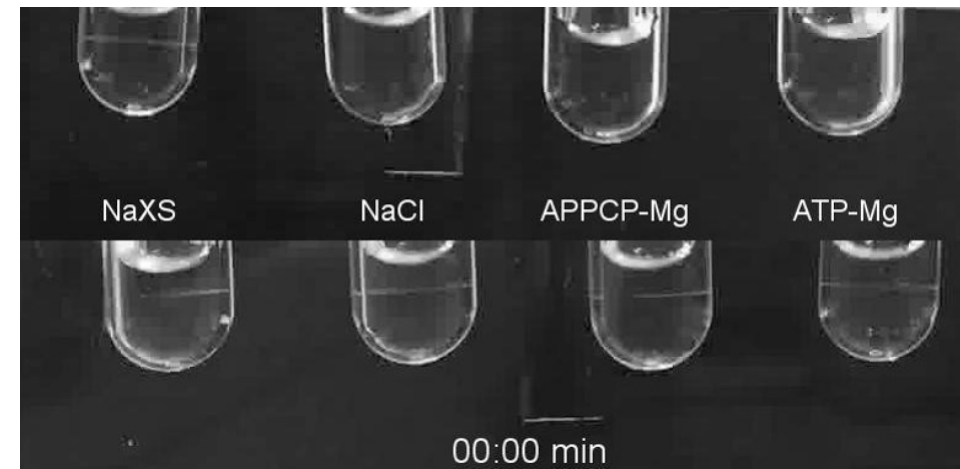
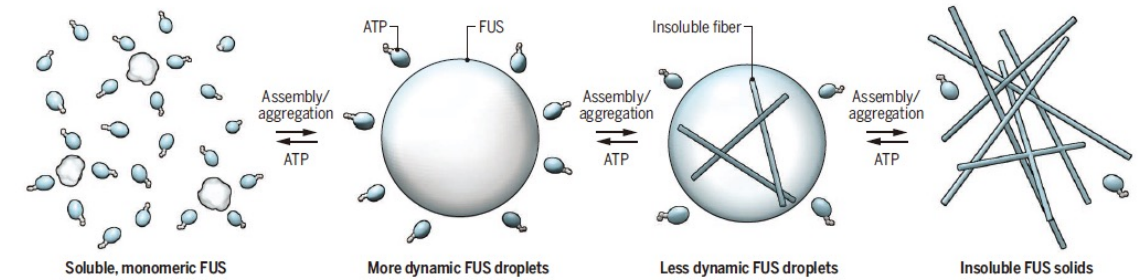
ATP distributions on the Protein Surfaces



A hidden function of ATP in the cells (hydrotrope in crowded conditions)

ATP controls solubility

K_m values for ATP-driven cell processes typically lie between 10 and 500 μM , whereas hydrotrope activity requires 2 to 8 mM of ATP. Cells maintain millimolar concentrations of ATP, perhaps to keep proteins (such as FUS) soluble by exploiting ATP as a biological hydrotrope.



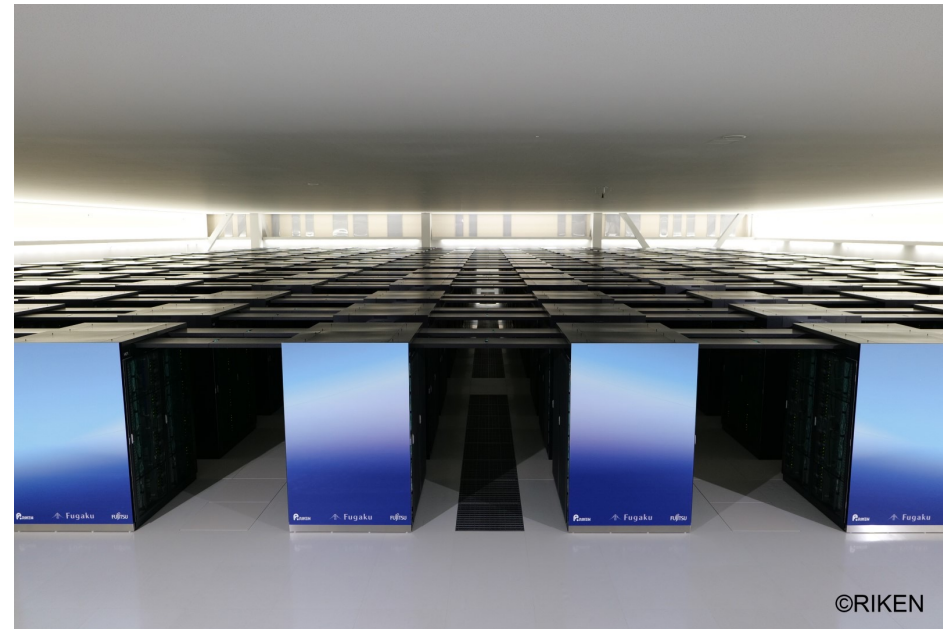
Interactions between proteins and ATPs are important.

Yu et al. *eLife* 5, e19274 (2016).

Patel et al. *Science* 356, 753-756 (2017)

The Supercomputer Fugaku

Node	CPU	Armv8.2-A SVE 512bit
	Performance	3.072TF (DP) 6.144TF (SP) 48 (+2) core
	Memory	32 GB
Number of Nodes per Rack		384 node x 396 rack 192 node x 36 rack
Total Number of Nodes		158,976
Network		Tofu Interconnect D
Peak Performance		488 PF (DP) 977 PF (SP)
Total Memory		4.85 PB
File IO		LLIO
OS		Red Hat Enterprise Linux 8



Top500	No. 1 in June and Nov. 2020
Graph500	No. 1 in June and Nov. 2020
HPL-AI	No. 1 in June and Nov. 2020
HPCG	No. 1 in June and Nov. 2020

Fugaku is now the fourth fastest supercomputer in the world.

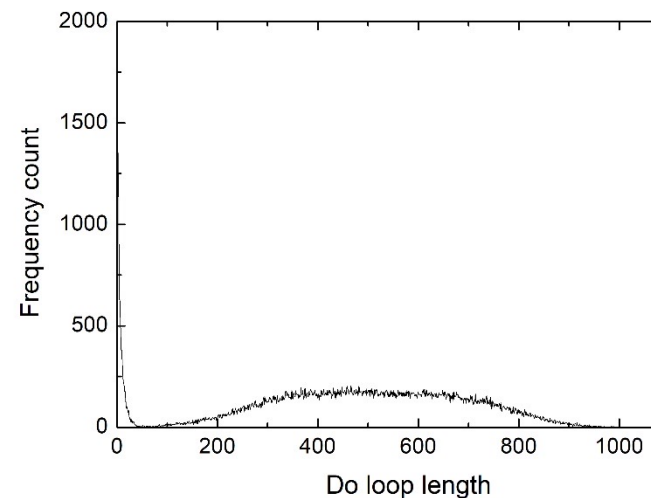
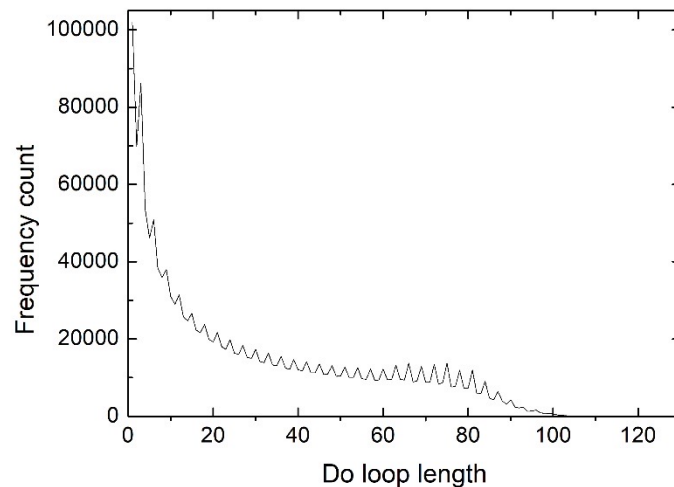
Optimization of GENESIS on Fugaku

New non-bonded interaction algorithm in GENESIS

```
do ijcel = 1, cell_pair
  obtain icel and jcel
  do i=1,natom(icel)
    do k=1,neighbor(i,ijcel)
      j = list(k,ijcel)
      interaction end do
    end do
  end do
end do
```



```
do icel = 1, cell
  do i=1,natom(icel)
    do k=1,neighbor(i,icel)
      list(k,i,icel)
      interaction end do
    end do
  end do
end do
```

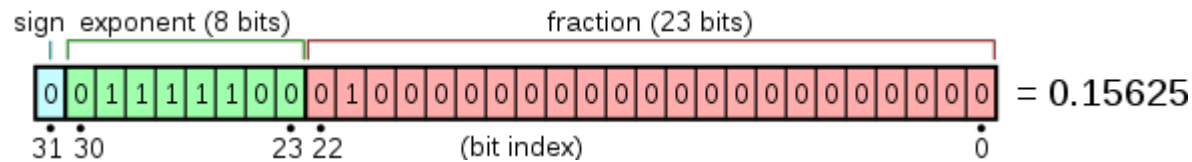


ApoA1 on one MPI processor

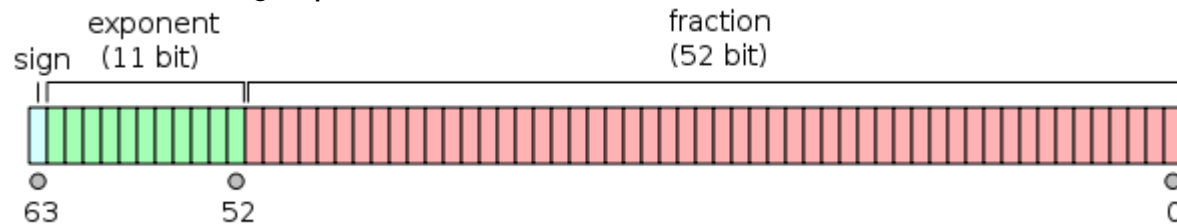


Single/Mixed/Double precision

- Single precision
 - 32 bit
 - Fast evaluation is possible by SIMD.
 - Less accurate than double precision



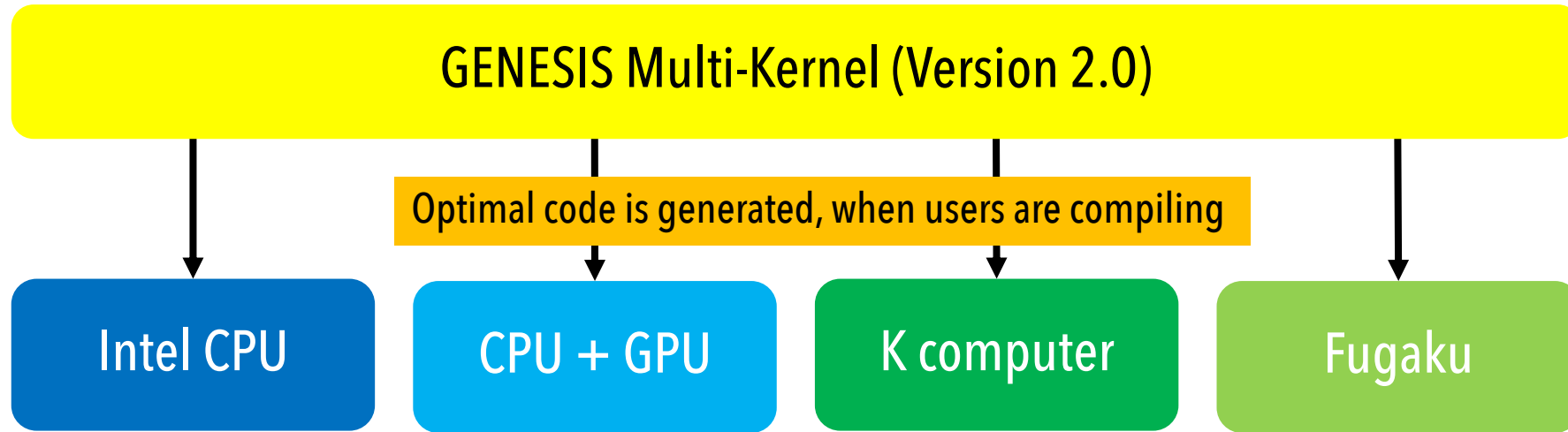
- Double precision
 - 64 bit
 - More accurate than single precision



- We can define precision levels in GENESIS 2.0 according to our purposes
 - Single precision: All real numbers except energy/constraints are described by single precision
 - Mixed precision: **Integration** by **double precision** and **force calculation** by **single precision**
 - Double precision: All numbers are described by double precision

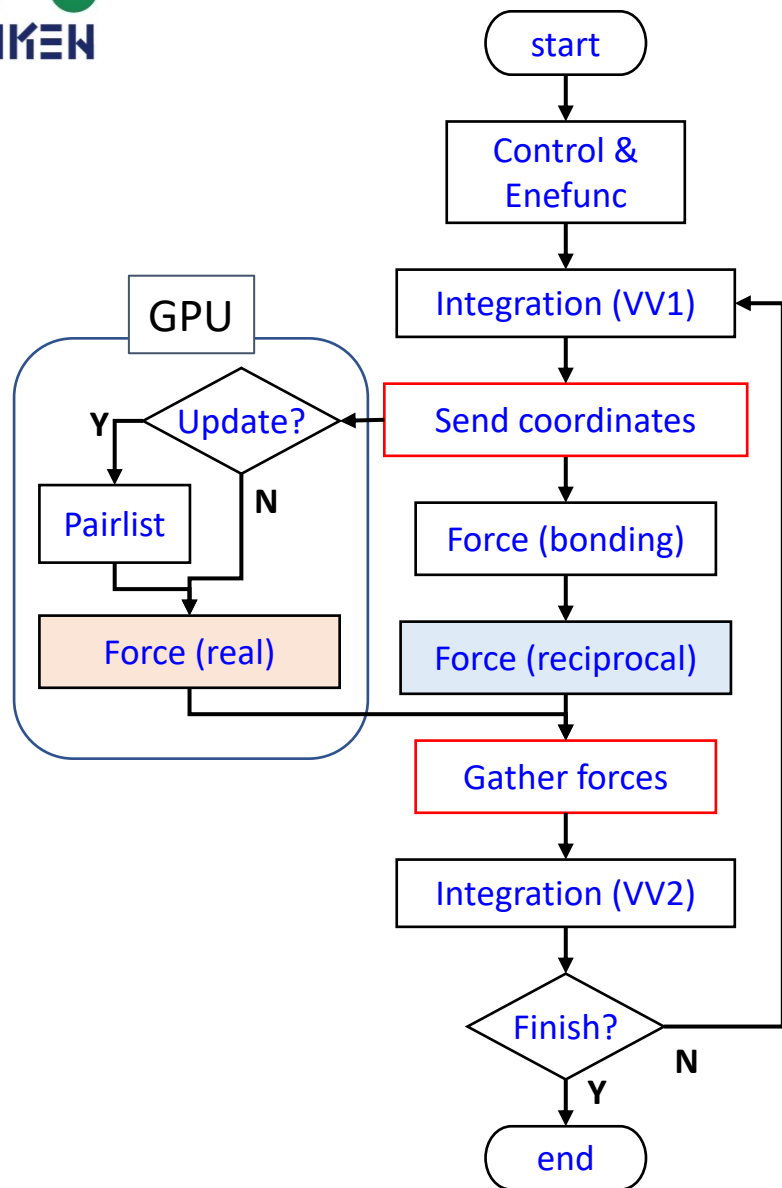
Multiple kernels for nonbonded energy calculations

User Friendly Multiple Kernels for Nonbonded Energy Calculations



- GENESIS 2.0 (the latest version) includes multiple kernels for nonbonded energy calculations.
- Users need to select a proper option when they compile GENESIS on each computational platform.
- Then, optimal code is generated automatically.
- Precision is also needed to select when compile GENESIS on each platform. We recommend `single_precision` for intel compiler and `mixed_precision` for Fugaku, considering the performance as well as accuracy.

Hybrid GPU+CPU application in GENESIS



1. Computation intensive work : GPU

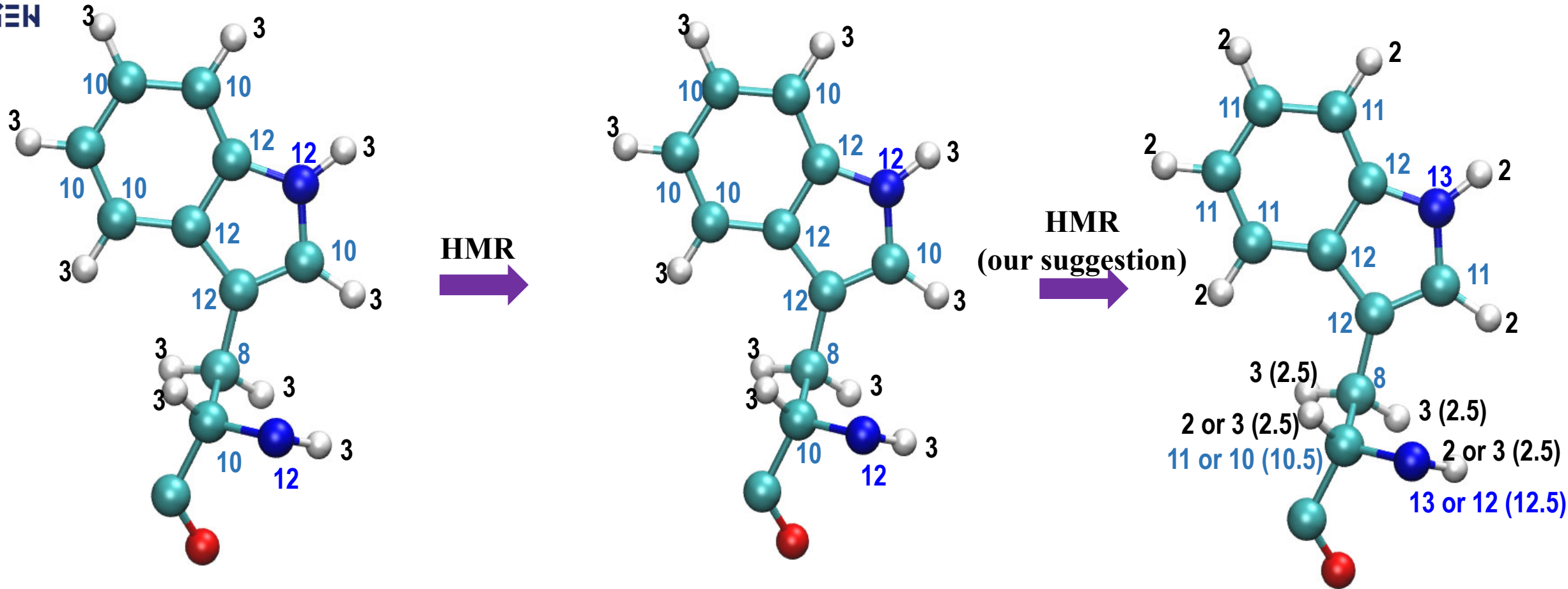
- Pairlist
- Real space non-bonded interaction

2. Communication intensive work : CPU

- Reciprocal space non-bonded interaction with FFT
- Bonded interaction
- Exclusion list

3. Integration is performed on CPU due to file I/O.

Our suggestion of HMR ratio



1. HMR ratio is 2 for XH_1 type or ring case
2. For XH_2 or XH_3 , we assign HMR ratio 3 (CHARMM) or 2.5 (AMBER)
3. It should be noted that this does not confirm the reliability of 5 fs all the time. To avoid shake error all the time, 3.5 fs with RESPA MTS could be better solution.

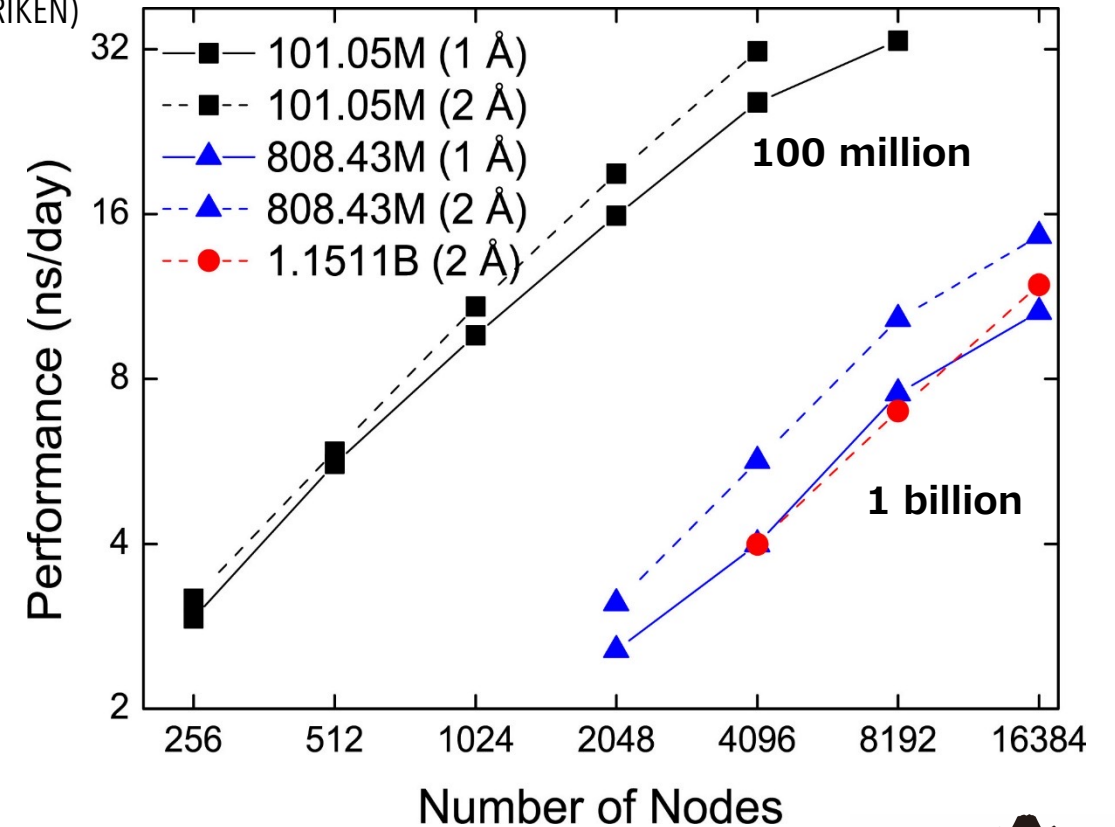
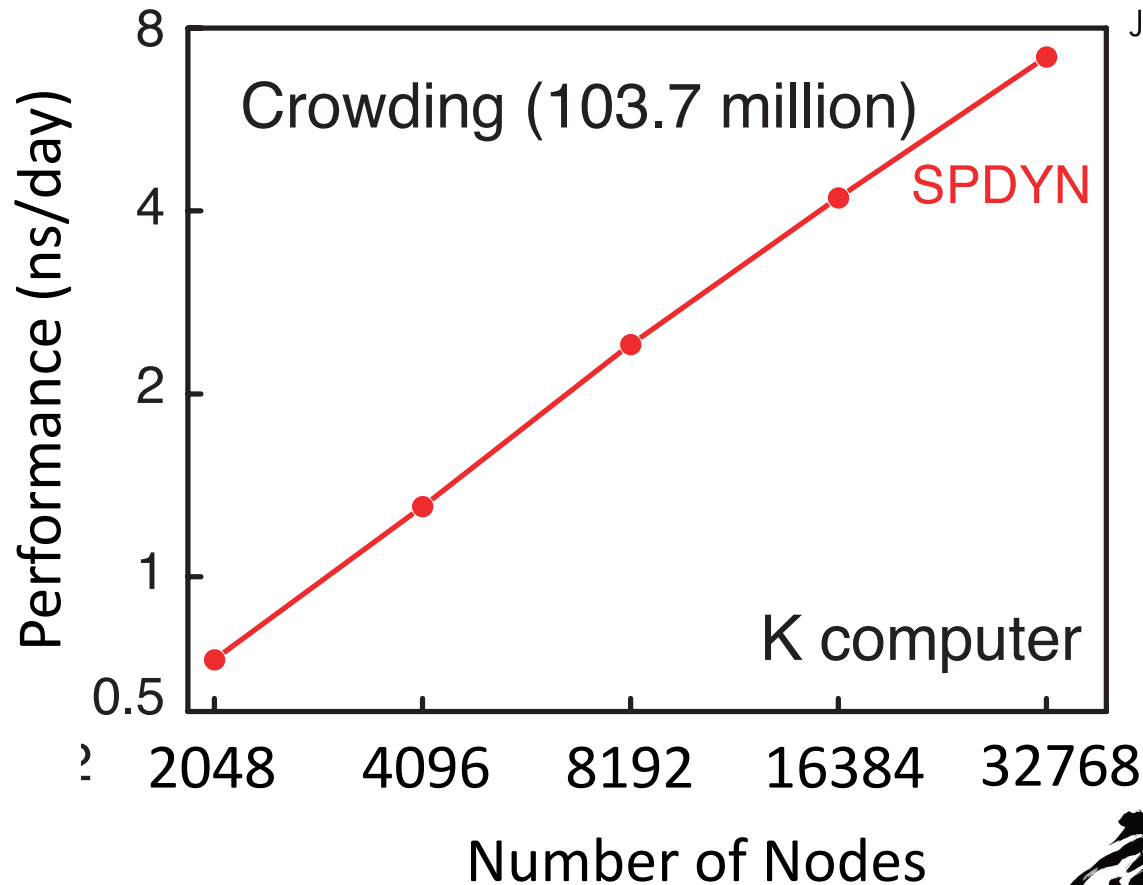
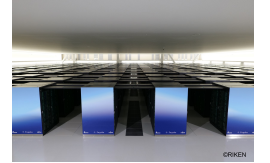
Acceleration of our MD software, GENESIS

GENESIS version 1 on K



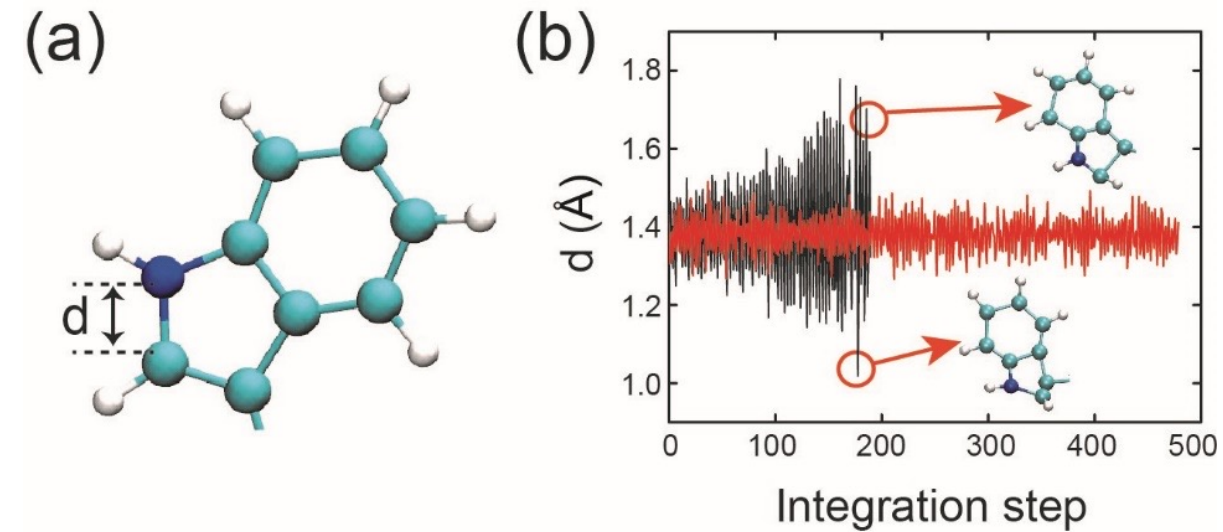
J. Jung (RIKEN)

GENESIS version 2 on Fugaku



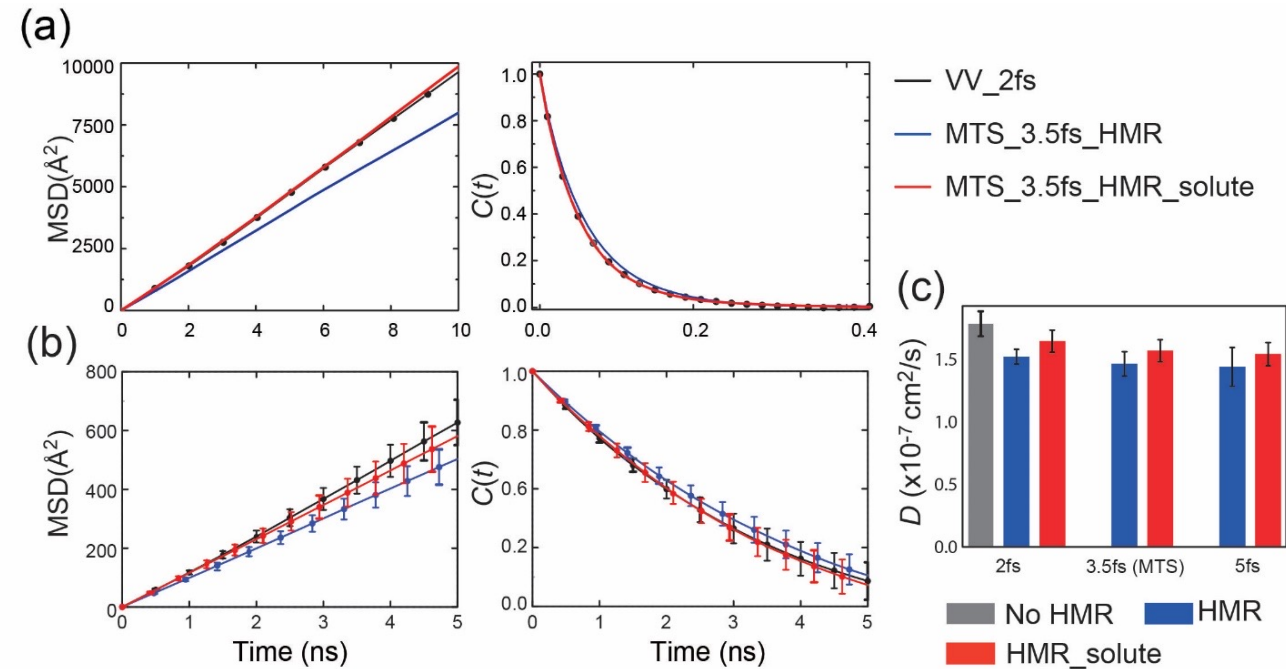
How accurate MD simulation with HMR?

Stable integration with a large time step



MD trajectories of a bond distance between nitrogen and carbon atoms in the pyrrole ring (“ d ” shown in Fig. 4(a)) of the ligand binding domain of AMPA receptor. The black line starts from 210 steps before the constraint error using HMR. In contrast, HMR_xh1 gives a stable trajectory of the distance, starting from the same structure. (Inset) Two structures of distorted indole rings.

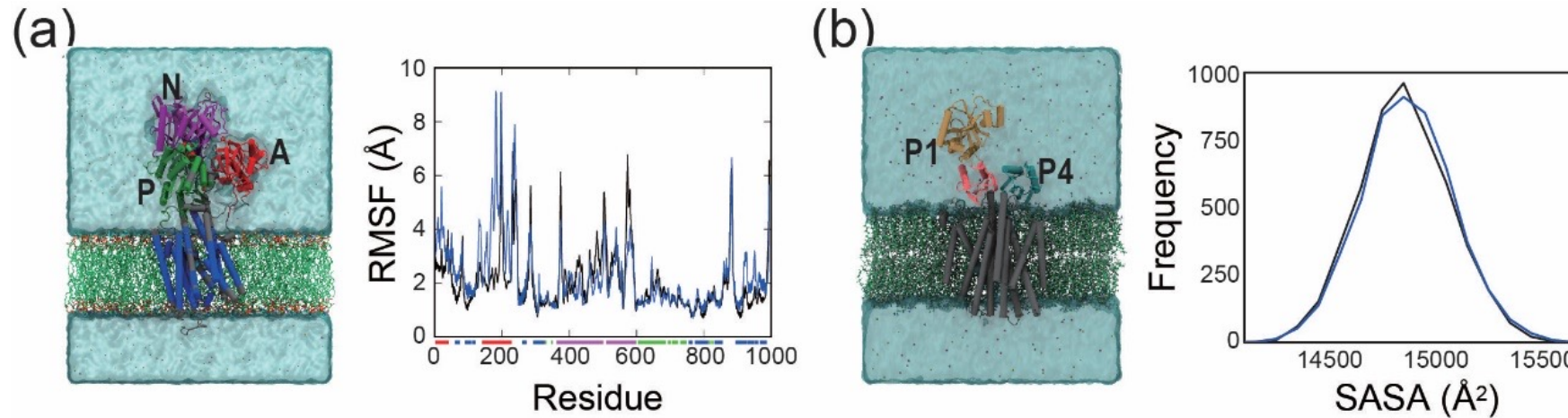
Diffusive motions of proteins and ligands



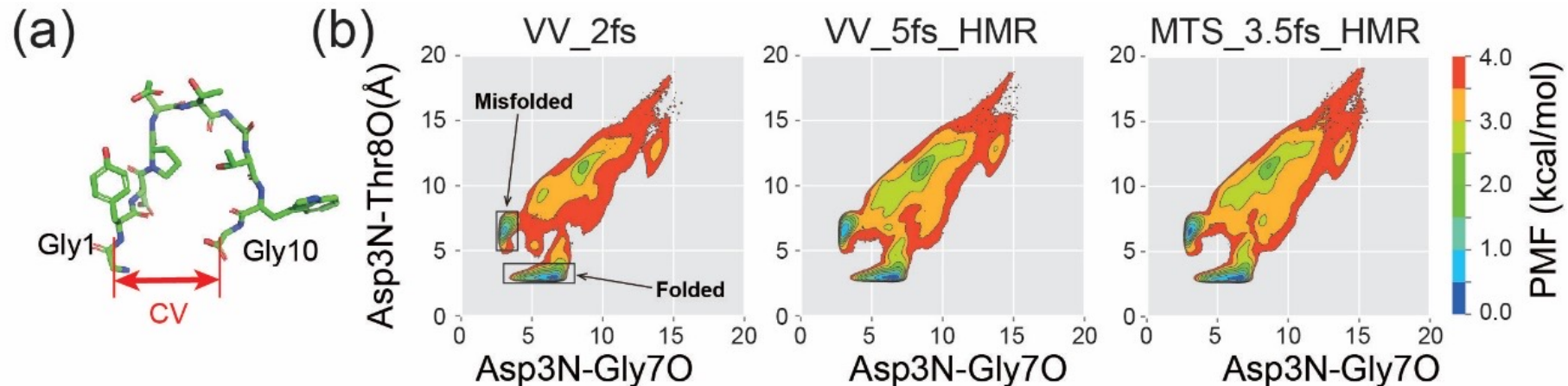
Translational mean squared displacements (left) and rotational correlational function (right) of (a) PP1 and (b) CI2. (c) Lateral diffusion coefficients of DPPC in 160DPPC systems in seven simulations.

How accurate MD simulation with HMR?

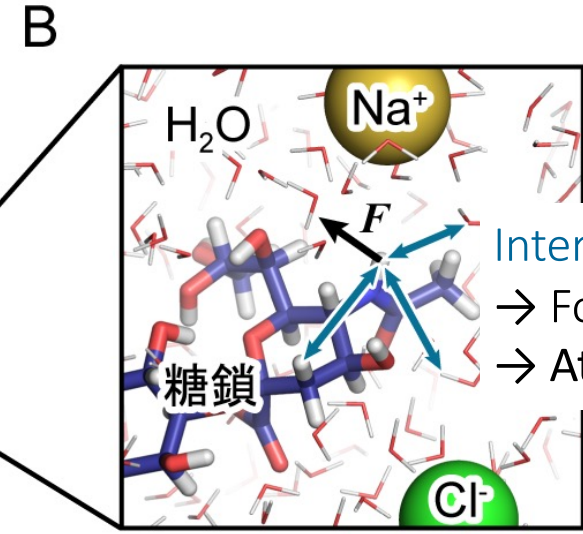
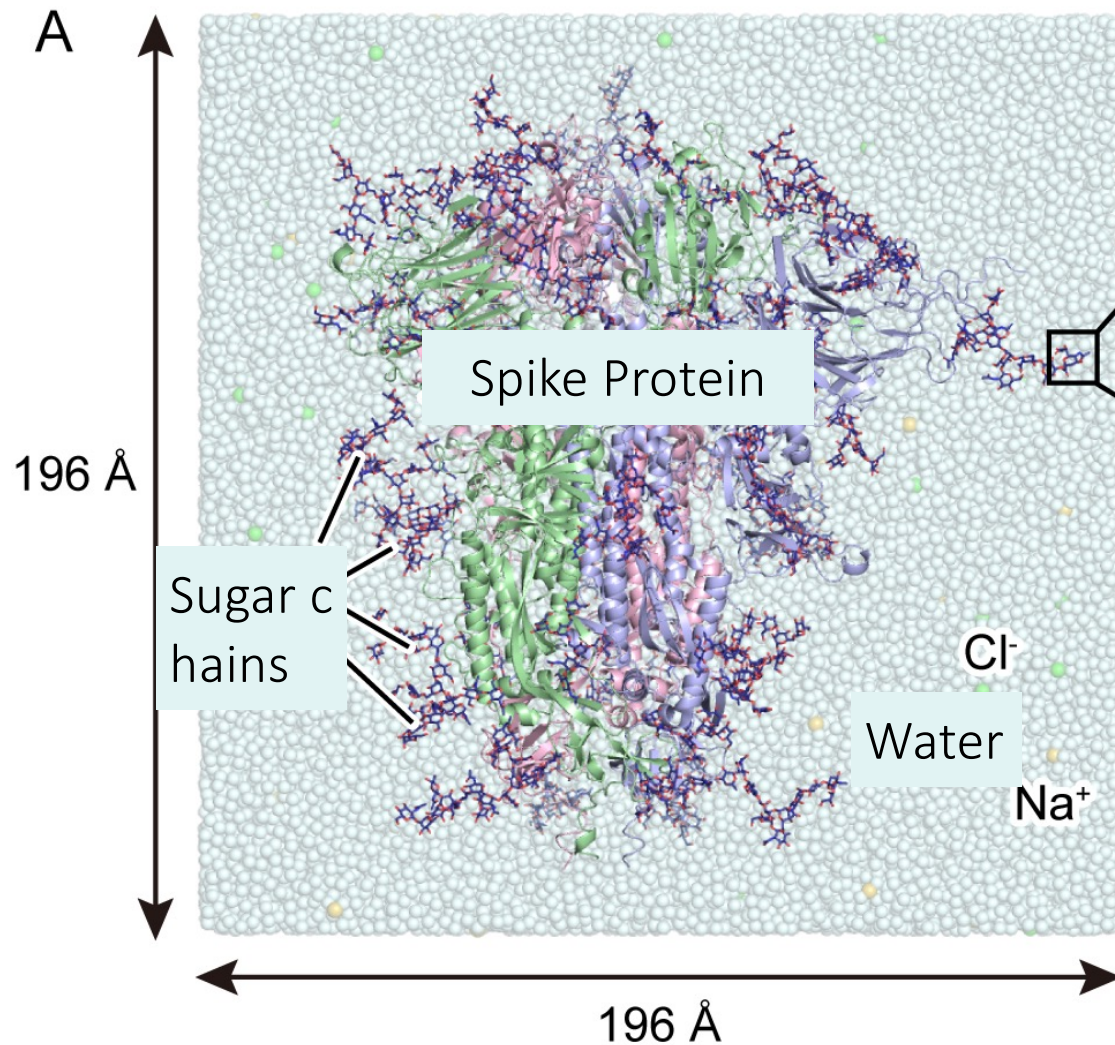
Membrane protein dynamics (a: Calcium pump, b: Sec translocon)



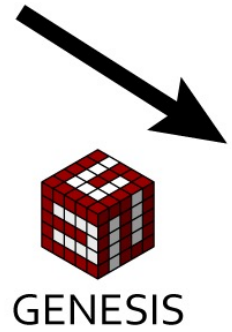
Free-energy landscape of Chignolin in water



MD simulations of SARS-CoV-2 S-protein on Fugaku



Interactions between atoms
→ Force (F) is applied to atoms
→ Atoms move

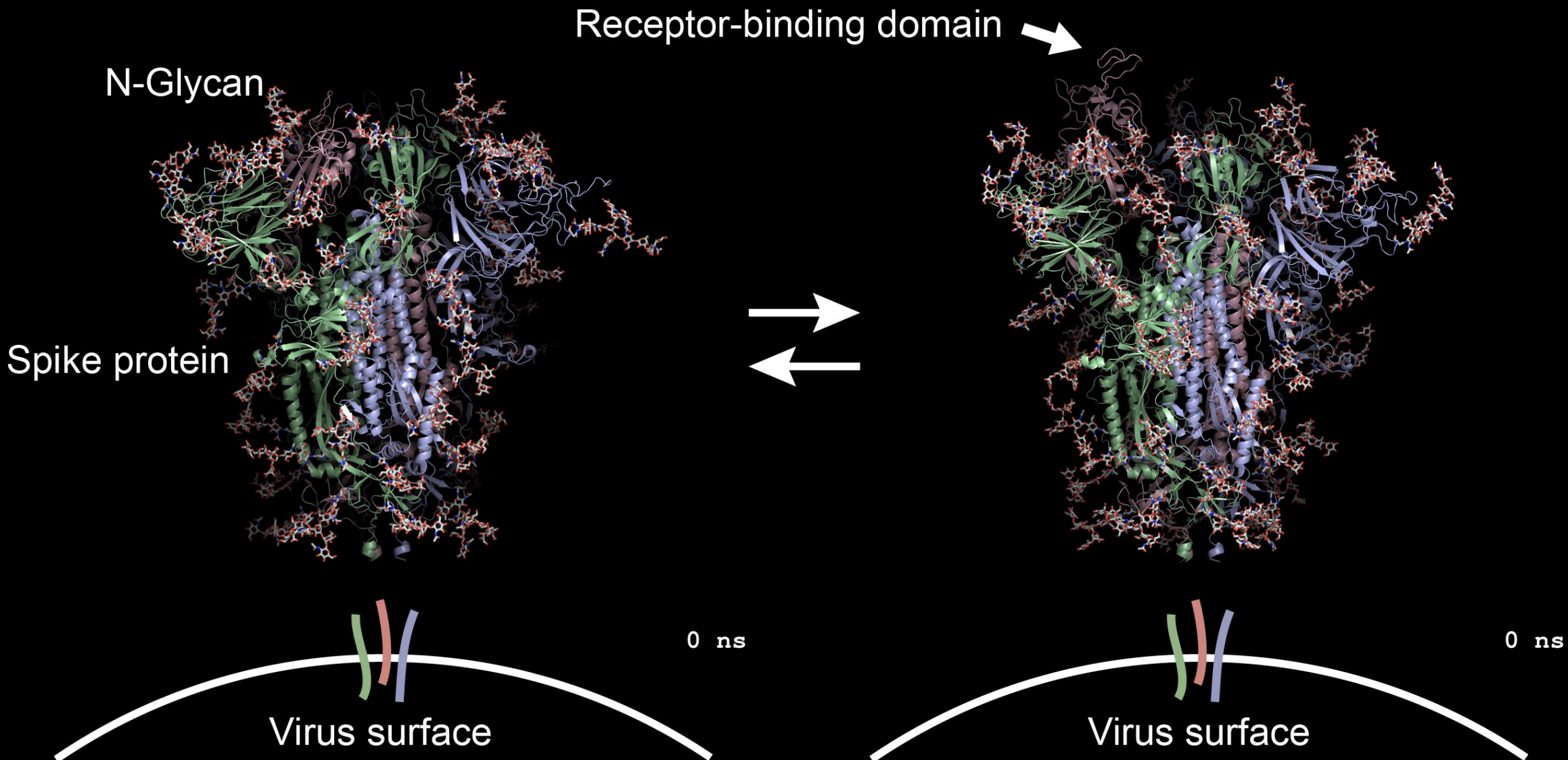


Simulation on Fugaku



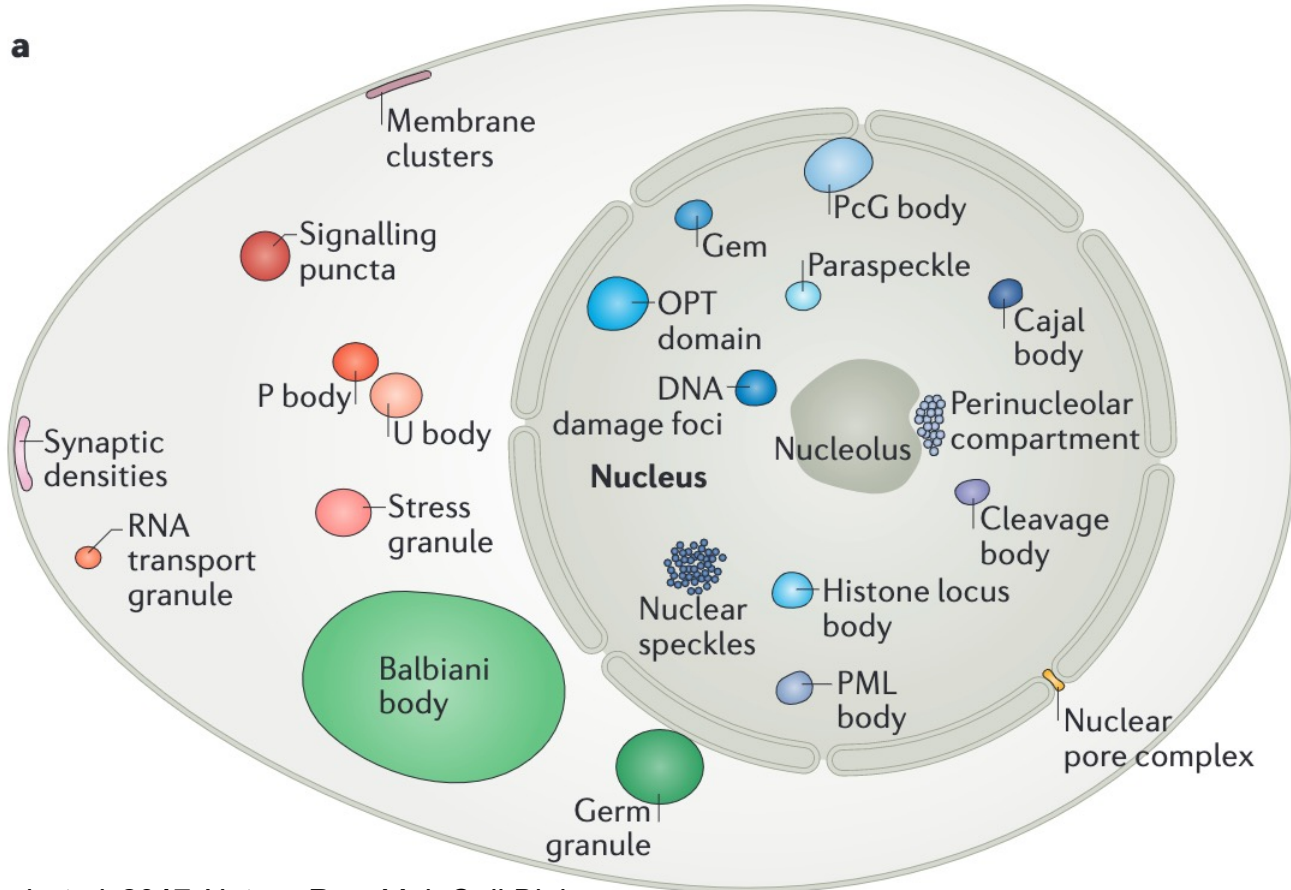
Down form

Up form

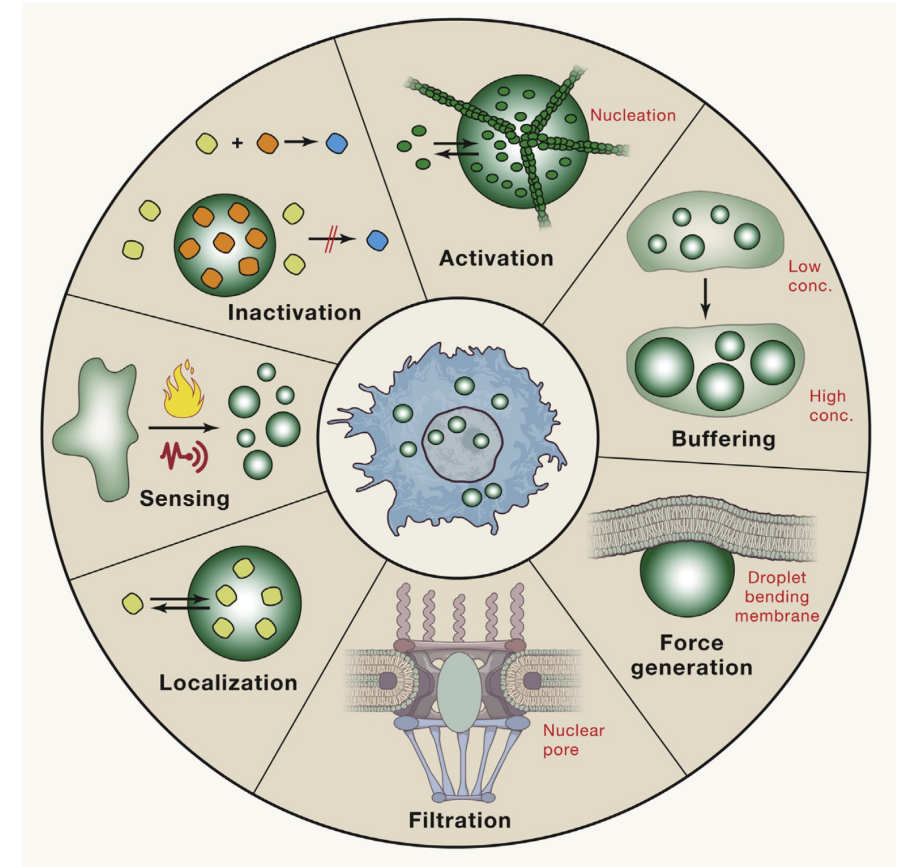


**What can we study in biology
(using a fast MD program on
supercomputers)?**

Liquid-liquid phase separation (LLPS)



Banani et al. 2017 *Nature Rev. Mol. Cell Biol.*



Alberti et al. 2019 *Cell*

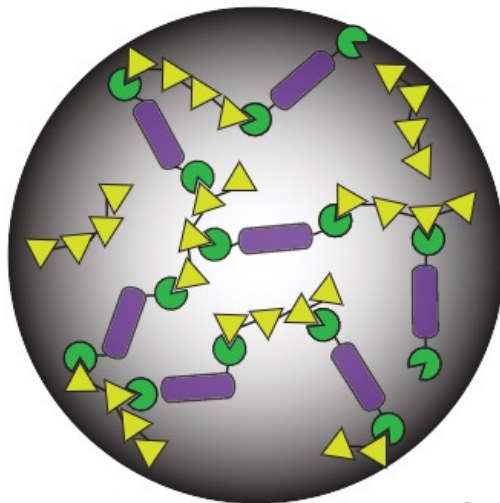
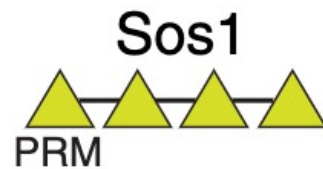
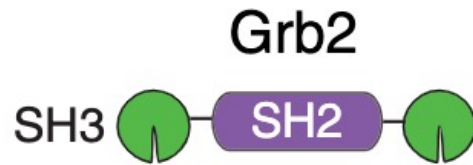
Names of LLPS in biology:

- Biomolecular Condensates
- Membraneless Organelles
- Liquid Droplets

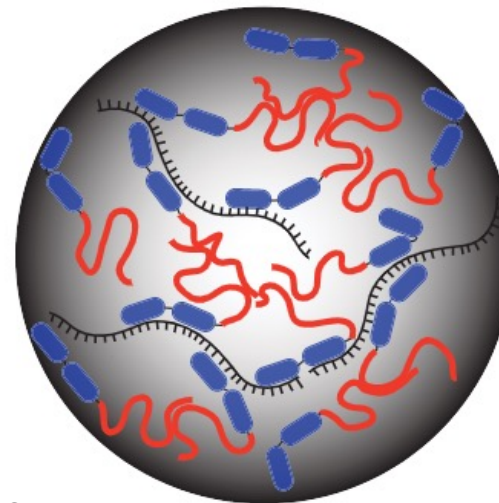
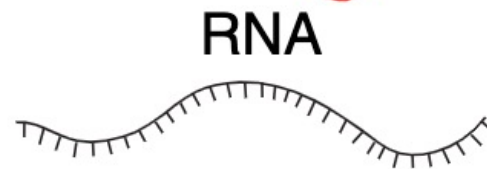
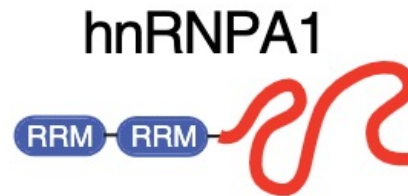
Multivalency: the General Principle of LLPS

Weak transient interactions between multivalent proteins drive LLPS.

A Multivalent Proteins

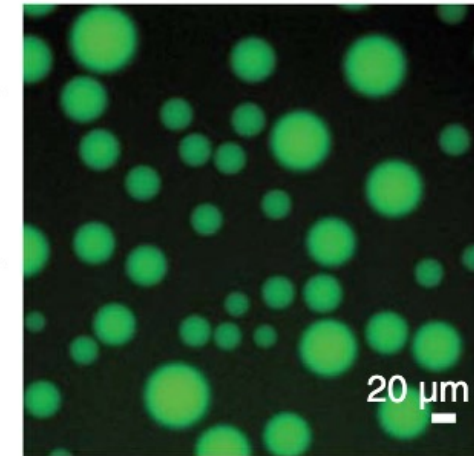


Disordered Proteins

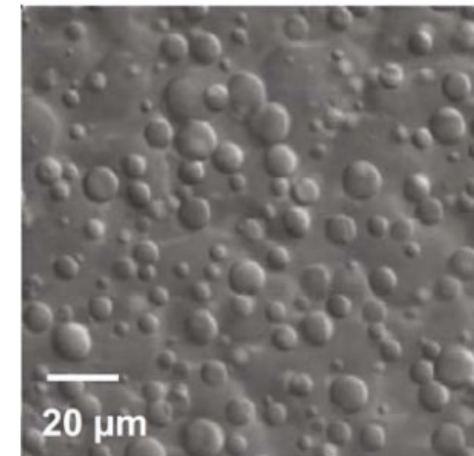


B

SH3₄+PRM₄

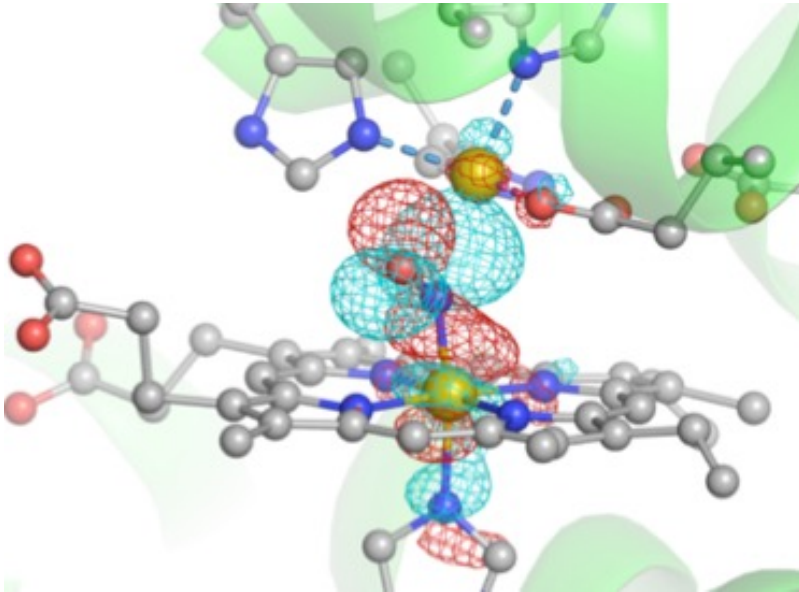


hnRNPA1



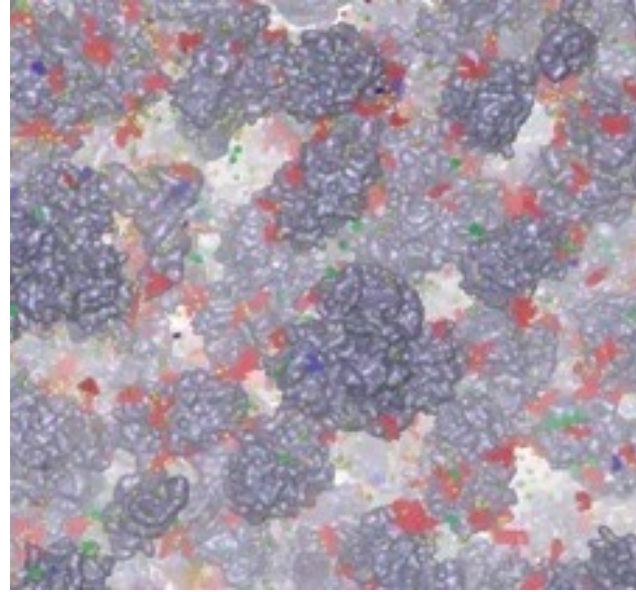
Multi-scale molecular models in GENESIS

Hybrid QM/MM MD ↔ Atomistic MD ↔ Coarse-Grained MD



Yagi et al., *JCTC* (2019)

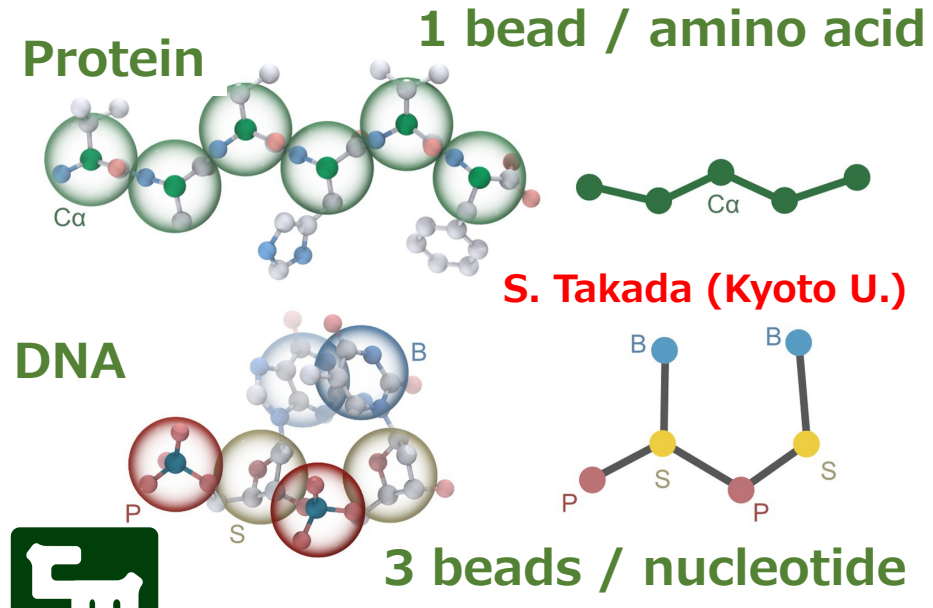
- Electronic structures
- Enzyme reactions
- Material designs
- Vibration and spectroscopy



Yu et al., *eLife* (2016)

- Membrane protein
- Protein/DNA complex
- Cellular environments

M. Feig (Michigan State U.)



S. Takada (Kyoto U.)

Tan et al. *PLoS Comp. Biol.* (2022)



- Slow dynamics
- Macromolecular interaction
- Data-driven simulations
- Cellular-scale simulations

Structure-based Coarse-Grained Models for Biomolecules

Residue-level coarse-graining: ~10 atoms / CG particle

- Protein: AICG2+**

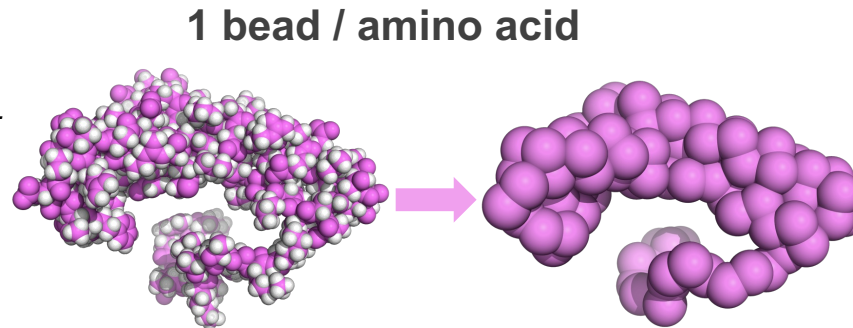
W. Li *et al.* 2014, *PNAS*.

- DNA: 3SPN.2C**

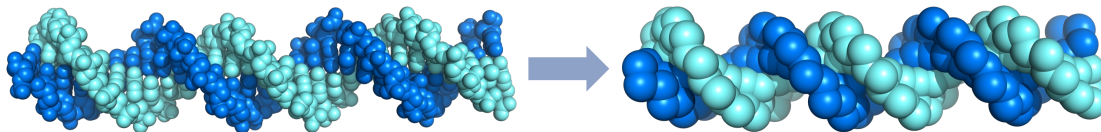
G. Freeman *et al.* 2014, *JCP*.

- RNA: Go-like**

N. Hori *et al.* 2012, *JCTC*.

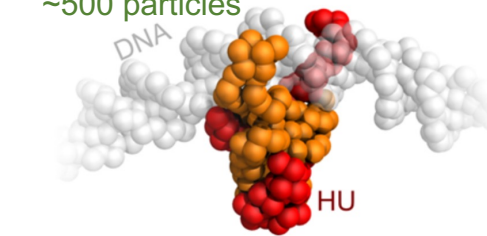


3 beads / nucleotide: Phosphate, Sugar, Base

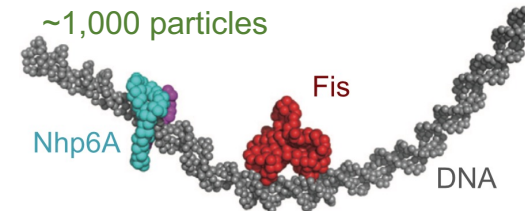


Residue-level CG models: protein-DNA systems

~500 particles



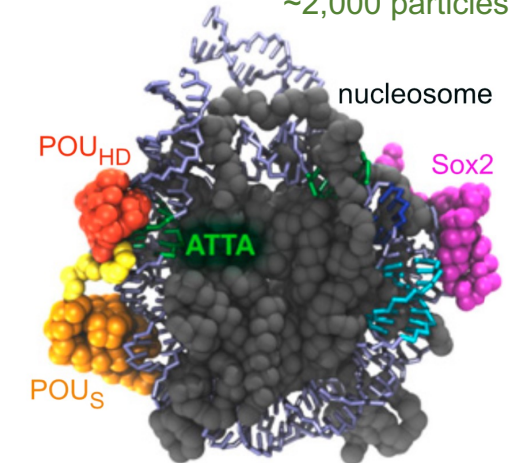
~1,000 particles



C.Tan *et al.* 2016, *JACS*.

K. Kamagata *et al.* 2020, *NAR*; 2021, *NAR*.

~2,000 particles



C.Tan *et al.* 2018, *JCTC*.

C.Tan *et al.* 2020, *PNAS*.

Protein-DNA: PWMcos

C.Tan *et al.* 2018, *JCTC*.



<https://www.cafemol.org/>



Prof. S. Takada
(Kyoto U.)



C. Tan
(Kyoto U. → RIKEN)

These CG models, which were originated from the structure-based Go-model, have been developed and used in **the CafeMol program by Prof. Shoji Takada's group.**

We (RIKEN) are collaborating with the Takada group (Kyoto U) to develop CG MD simulations in GENESIS software.

CG models in GENESIS MD software

GENESIS supports CG models of protein, RNA, DNA and Lipid



C. Tan (RIKEN)



D. Ugarte La Torre (RIKEN)

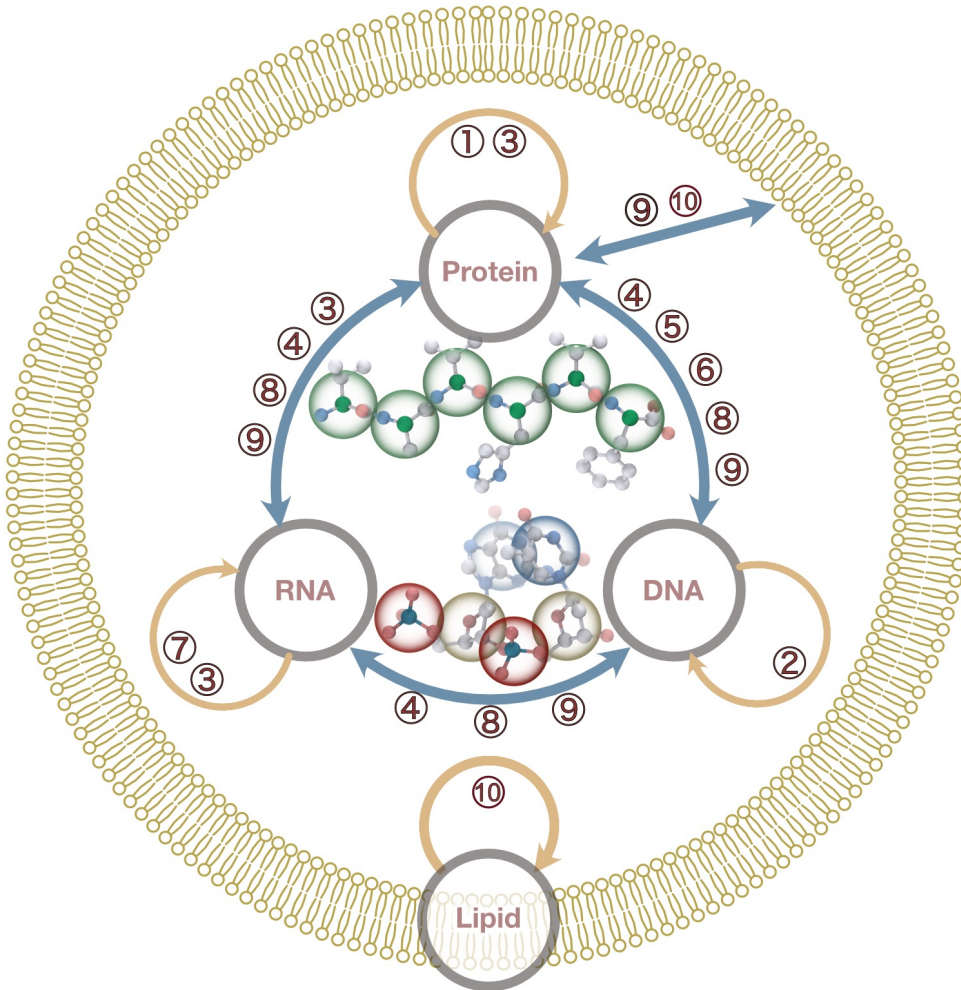


GENESIS

Generalized-ensemble simulation system

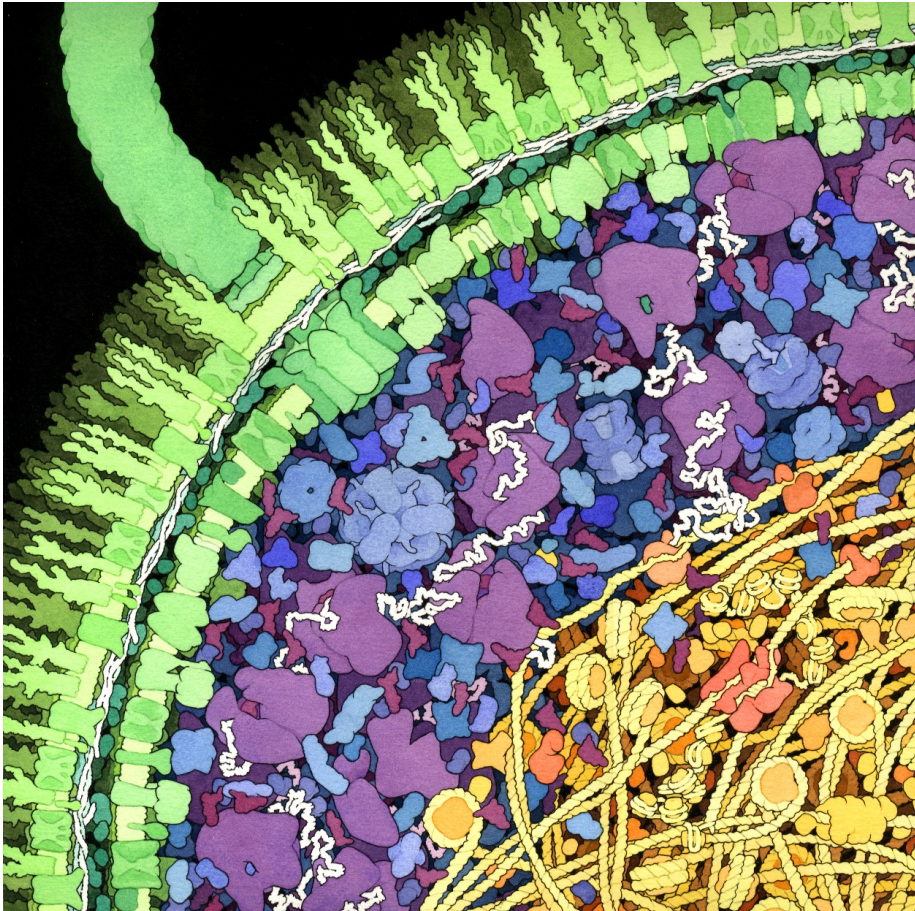
Residue-level coarse-grained models

1. W. Li *et al.* (2014). *Proc. Natl. Acad. Sci.*
2. G.S. Freeman *et al.* (2014). *J. Chem. Phys.*
3. G.L. Dignon *et al.* (2018). *PLoS Comput. Biol.*
4. C. Clementi *et al.* (2000). *J. Mol. Biol.*
5. C. Tan & S. Takada (2018). *J. Chem. Theory Comput.*
6. G.B. Brandani *et al.* (2018) *Nucl. Acids Res.*
7. N. Hori. & S. Takada (2012). *J. Chem. Theory Comput.*
8. C. Tan & S. Takada (2016). *J. Am. Chem. Soc.*
9. P. Debye & E. Hückel (1923). *Physikalische Zeitschrift*
10. D Ugarte La Torre *et al.* (2023). *J. Chem. Phys.* ← **New, for lipids!**



Summary and Perspectives

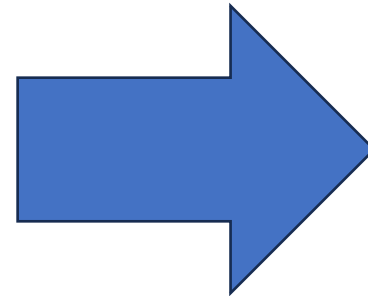
Macromolecular Crowding Effect



Zimmerman and Minton, *Ann. Rev. Biophys. Biomol. Struct.* (1993) 22: 27-65.

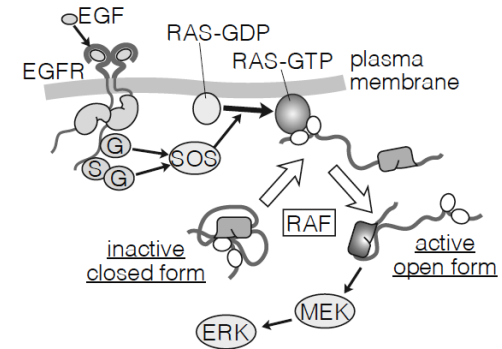
Zhou, H.X., Rivas, G., Minton, A.G., *Ann. Rev. Biophys.* (2008) 37, 375-397.

**Multi-scale models
(AA, CG, QM/MM)
Enhanced Sampling
(REMD, REUS, gREST,
GaMD, etc.)**



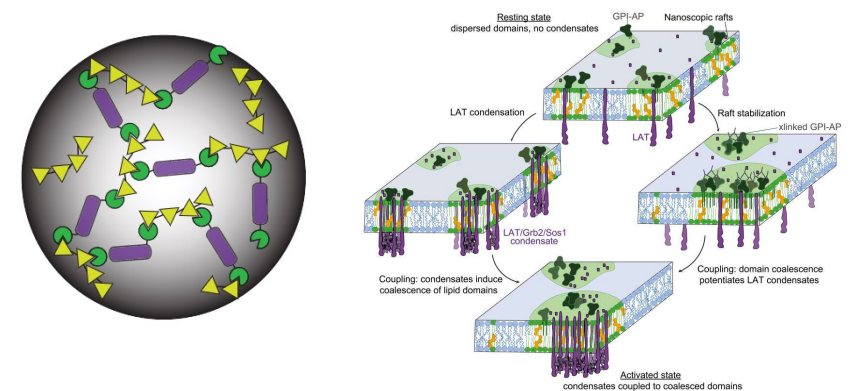
**Larger biological
systems in
atomistic or
residue-level MD
simulations**

Signal Transduction Pathway



Okamoto, Hibino, Sako. *BBA – General Subjects* 1864 (2020) 129358.

Protein Condensates



Shin et al., *Science* 357, 1253 (2017)

Wang et al., *Sci. Adv.* 9, eadf6205 (2023)

Acknowledgement

Collaborators

RIKEN:

- Dr. Jaewoon Jung (RIKEN CPR/R-CCS)
- Dr. Chigusa Kobayashi (RIKEN/R-CCS)
- Dr. Cheng Tan (RIKEN/R-CCS)
- Dr. Diego Ugarte La Torre (RIKEN/R-CCS)
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- Dr. Kiyoshi Yagi (RIKEN/CPR)
- Dr. Takaharu Mori (RIKEN/CPR→Tokyo U. Sci)

For Bacterial cytoplasm

- Prof. Michael Feig (Michigan State University)

For CafeMol

- Prof. Shoji Takada (Kyoto University)

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- MEXT "Priority Issue 1 on post-K computer"
- MEXT "Program for Promoting Researches on the Supercomputer Fugaku"
- MEXT Grant-in-Aid for Scientific Research (S) (19H05645)
- MEXT Grant-in-Aid for Transformative Research Area (A), Cross-Scale Biology (21H05249)
- RIKEN Pioneering Projects (Dynamic Structural Biology/Glycolipidologue Initiative/Biology of Intracellular Environments)
- Research grants from RIKEN R-CCS and BDR

Computer resources:

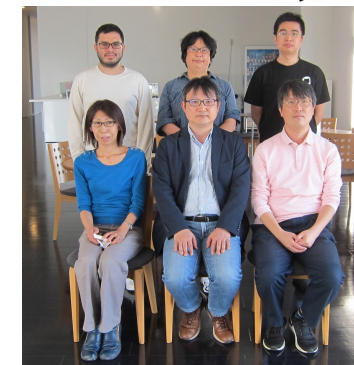
- RIKEN HOKUSAI GreatWave, HOKUSAI BigWaterfall, K computer, and Fugaku supercomputer
- HPCI supercomputer resources (hp170254, hp180201, hp180274, hp190097, hp190181, hp200129, hp200135, hp210107, hp210177, hp220087).

RIKEN Sugita Lab

BDR @ Kobe (2022)



R-CCS @ Kobe (2022)



CPR @ Wako (2023)

